

Luminex Based Assay Development

Stephen Angeloni, Ph.D. Sr. Field Application Scientist Luminex Corporation

Luminex Technology A Broad Range of Applications

Research **Human Diagnostics** Animal Health Ag Sciences **Food Safety Biodefense Environmental Monitoring**





Escondido's

Luminex Corporate Offices Global Presence





Luminex

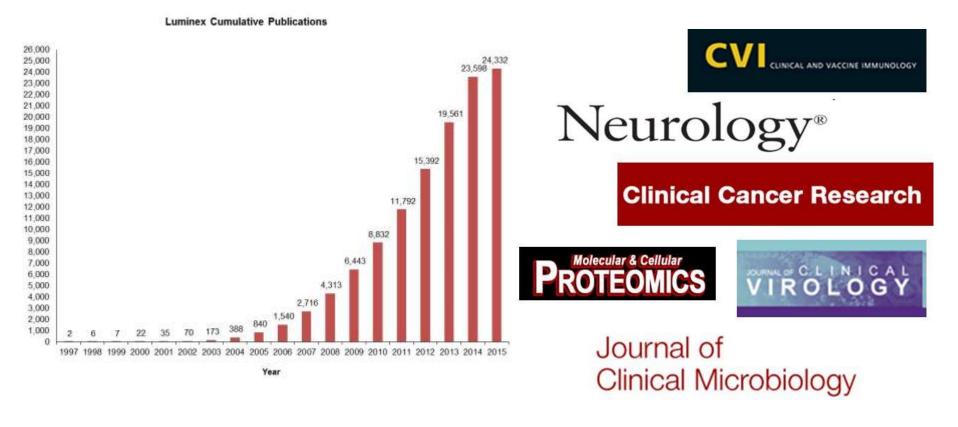
Global Presence: Over 11,000 instruments shipped globally





Luminex **Propelling Knowledge**

Over 24,000 peer-reviewed scientific studies and growing For complete listing see www.luminexcorp.com/publications



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Luminex Partners

Over 60 partners worldwide



Luminex Diagnostics More than 60 US FDA 510(k) Cleared Platforms











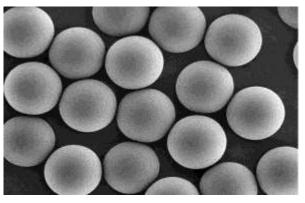


xMAP Technology: An Overview

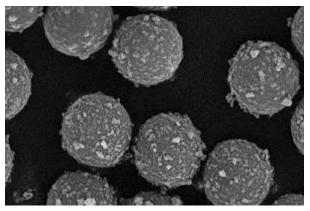
xMAP Technology

Microsphere-based Arrays

MicroPlex™ Microspheres 5.6 um



MagPlex[™] Superparamagnetic microspheres 6.5 um



Microsphere Advantages

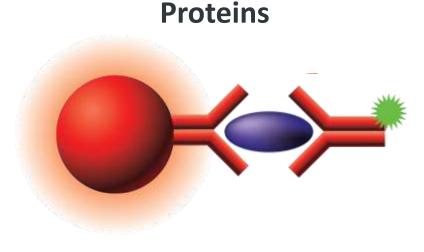
- Fast liquid phase kinetics
- Proprietary dyeing process
- Unique bead signatures
- High surface-to-volume ratio
- Surface carboxyl groups



xMAP Applications

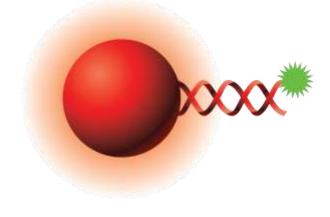
An Incredibly Flexible Platform

Bead sets can be coated with reagents specific to many bioassays:



- Immunoassays » Sandwich Capture » Multiplex ELISA
- Receptor-Ligand/Protein interaction Assays
- Enzyme Substrate

Nucleic Acids

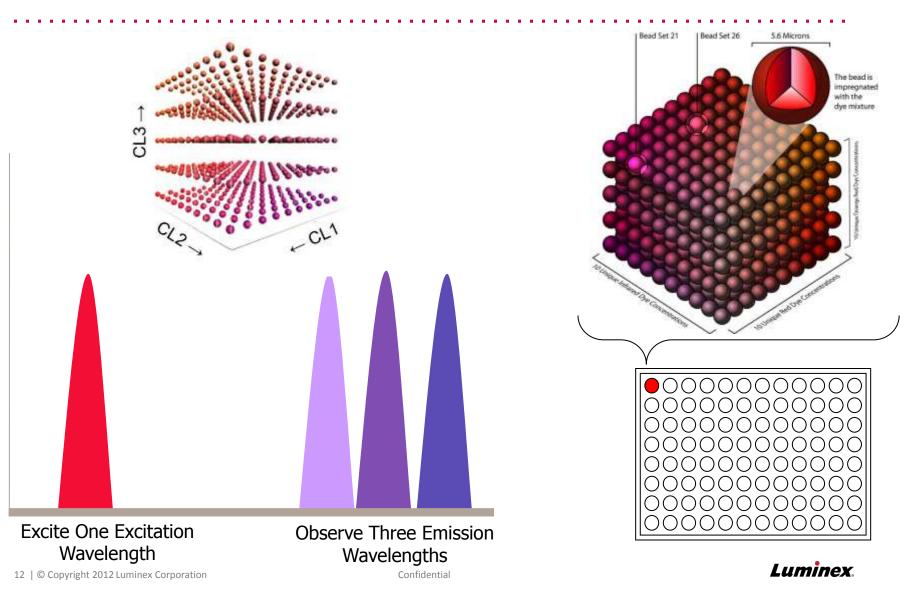


- Gene ExpressionmicroRNA Profiling
- Genotyping
 - »SNP
 - »CNV
 - »Sequence Detection

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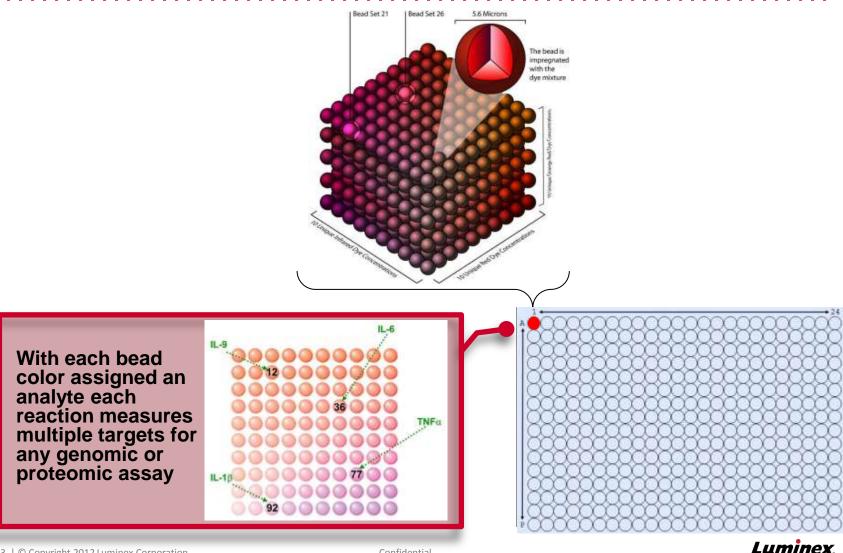
Spectral Addressing

Multiplexing to 500 Analytes/Well with MagPlex beads



Spectral Addressing = Multiplexing

Each Bead Associated with a Specific Analyte in each reaction



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Several Advantages to Multiplexing

Use less sample, less reagents, get more data faster, get more accurate data

Real Examples

A 107 plex Gene Expression assay.

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14 samples in triplicate. Done in 42 wells of a 96 well plate.

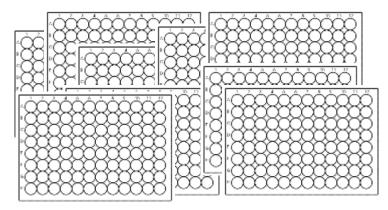
Single plex chemistry like TaqMan or another real time PCR chemistry would required 4,494 wells = 46.8 96 well plates.

24 plex Protein assay.

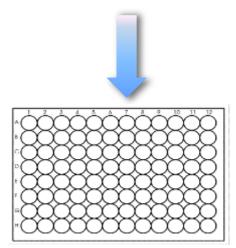
Our partners make protein assays that can analyze as many as 43 proteins or more with some kits. Running a 24 plex assay on 27 samples in triplicate required 81 wells of a 96 well plate.

A standard single plex ELISA chemistry would require 1,944 wells and more than 20 96 well plates. If 43 marker assay, single plex would need 3,483 reactions or 36.3 96 well plates

Single Plex Reactions



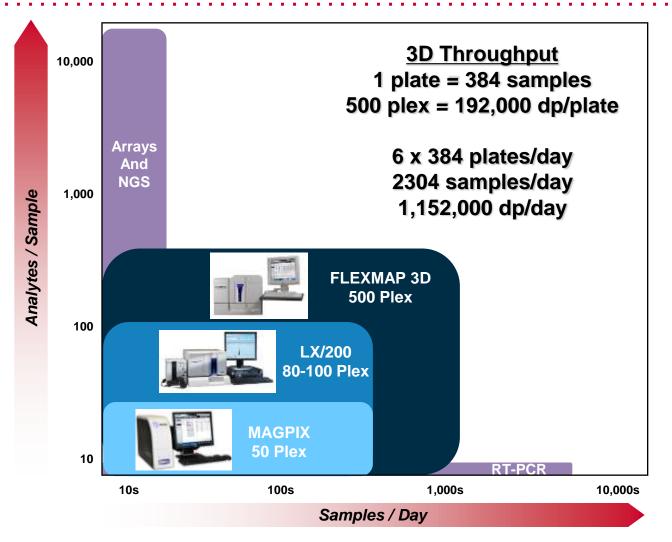
Luminex Multiplexing





Luminex Instruments

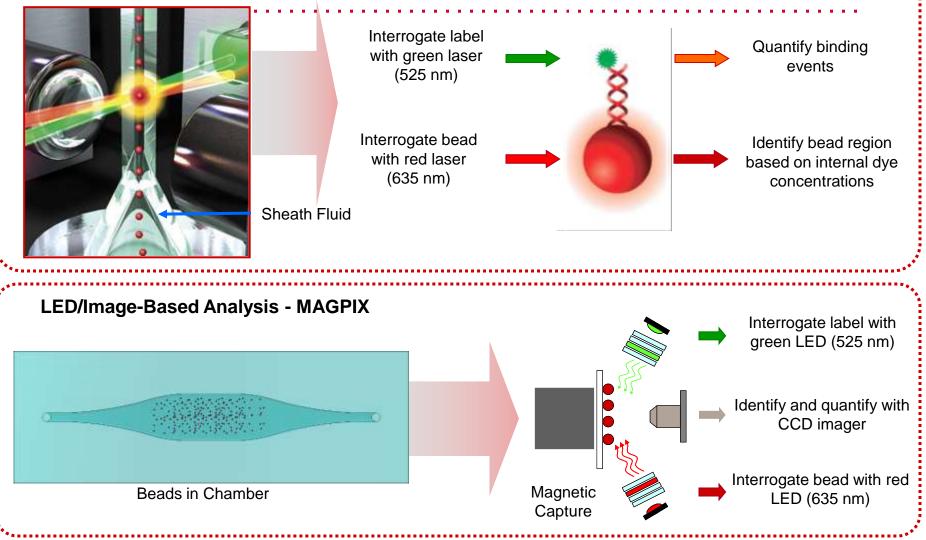
Unique Combination of Flexibility, Multiplexing and Throughput





Flow Cytometry and Imaging Comparison

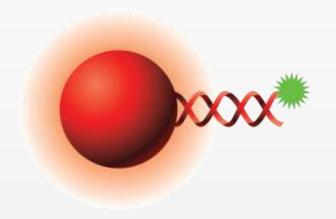
Flow Cytometry-Based Analysis – Luminex 200 & FLEXMAP 3D



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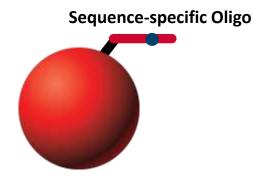


Genomic Assays



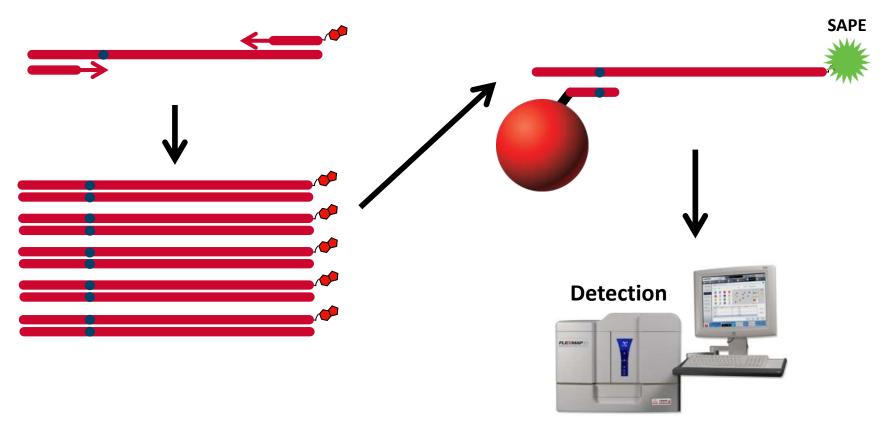


- Couple sequence-specific C18 spacer amine-modified oligo to bead
- Probes are matched for length ~20bp for SNPs, 50-70bp okay for unrelated sequences and mismatches are centered or distributed





- Label target nucleic acid sequence to be detected with biotin PCR primer
- Amplicons should be \leq 300bp if possible
- Denature, hybridize to beads, incubate with SAPE and detect



DDL/GSK HPV Genotyping Assay



A Human Papilloma Virus Testing Algorithm Comprising a Combination of the L1 Broad-Spectrum SPF₁₀ PCR Assay and a Novel E6 High-Risk Multiplex Type-Specific Genotyping PCR Assay

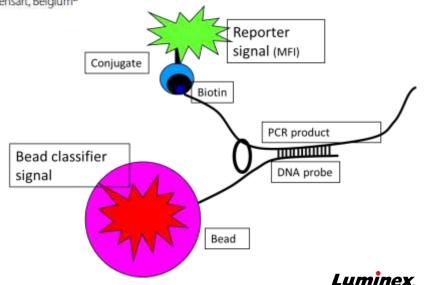
Dirk van Alewijk,^a Bernhard Kleter,^a Maarten Vent,^a Jean-Marc Delroisse,^b Maurits de Koning,^a Leen-Jan van Doorn,^a Wim Quint,^a Brigitte Colau^b

DDL Diagnostic Laboratory, Rijswijk, The Netherlands^a; GlaxoSmithKline Vaccines, Rixensart, Belgium^b



DDL DIAGNOSTIC LABORATORY

Partner in advanced diagnostic testing



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Luminex assays Developed and Published by DDL

DDL is a contract research organization and is available for collaborations and service

Target	Multiplex	Intended use	Reference
HPV (high-risk mucosal / anogenital)	21-plex	Research	Geraets et al, 2009
Chlamydia trachomatis	16-plex	Research & Epidemiology	Quint <i>et al,</i> 2009
HPV (wart)	25-plex	Research & Epidemiology	De Koning <i>et al,</i> 2010
HPV (high-risk mucosal / anogenital)	19-plex	Research & Epidemiology	Van Alewijk <i>et al,</i> 2013
HPV (mucosal / anogenital)	36-plex	Research & Epidemiology	Kleter <i>et al,</i> to be submitted



Partner in advanced diagnostic testing



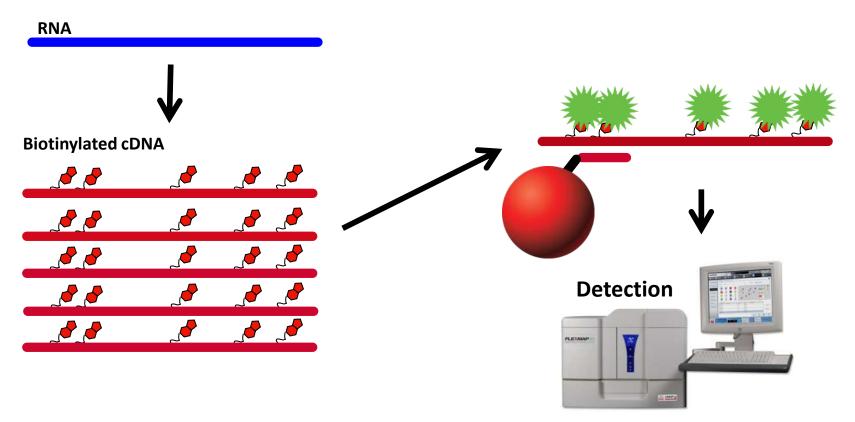
xMAP[®] Salmonella Serotyping Assay RUO Kit Training



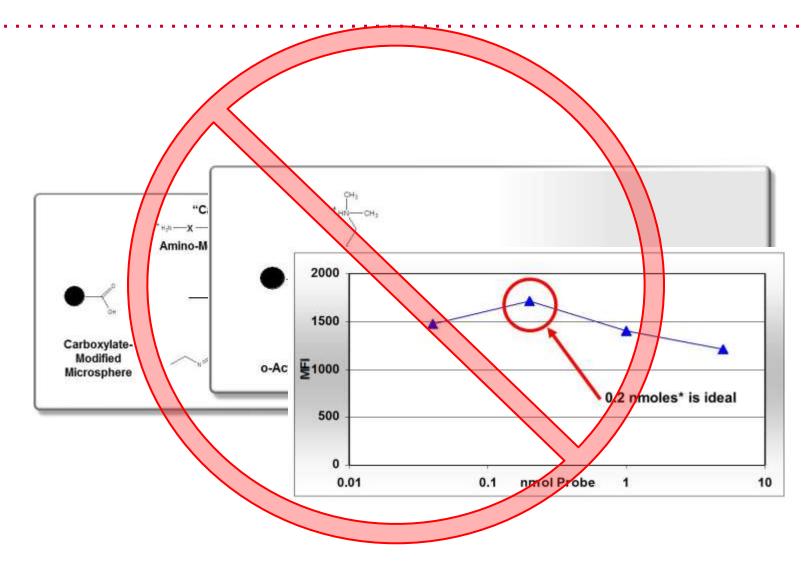


BADGE Gene Expression Assay

• BADGE, Beads Array for the Detection of Gene Expression, a highthroughput diagnostic bioassay. Yang, L., D.K. Tran, and X. Wang, *Genome Res*, 2001. 11(11): p. 1888-98.



Requires optimization of coupled capture oligonucleotides

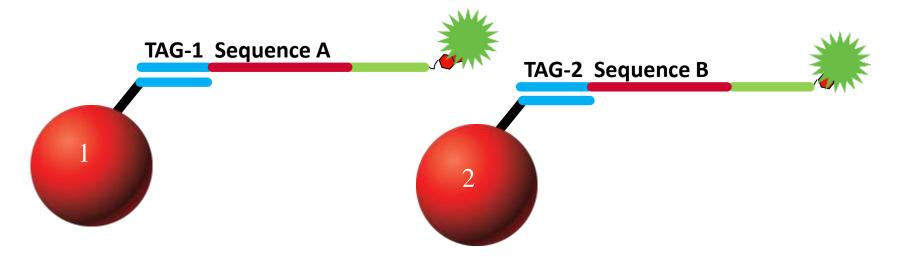


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xTAG Technology Simplifying Nucleic Acid Assays

Simplifying Nucleic Acia Assays

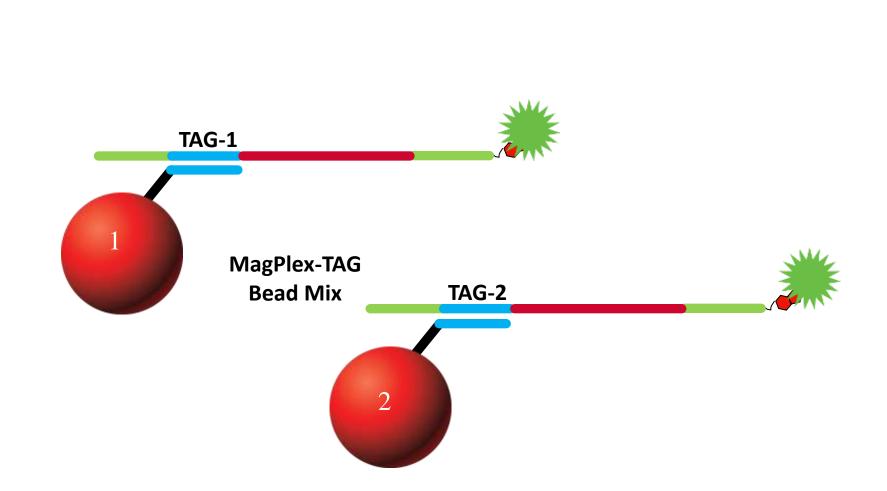
- Proprietary sets of 24 base oligonucleotides are coupled to xTAG beads
- Each xTAG bead region's tag is complementary to a specific tag sequence engineered onto reporter molecules
- This approach as been validated and used in multiplexed clinical genetic tests





Key to using xTAG Technology

For Simplifying Nucleic Acid Assays





Using xTAG Technology For Different types of Nucleic Acid Assays

- O Gene expression
- SNP typing, Specific sequence detection
- O Copy Number Variation Analysis (CNV)
- O miRNA analysis
- And variations of these chemistries for a number of applications



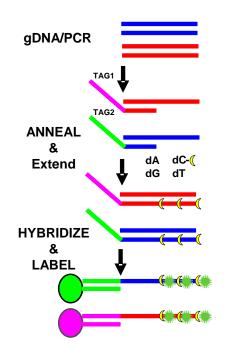
xTAG Chemistries

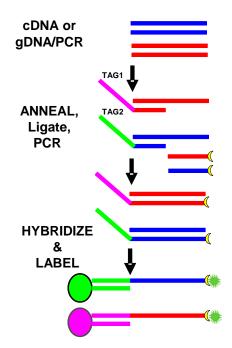
For Different DNA Analysis Assays

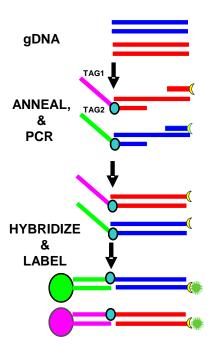
Allele Specific Primer Extension

LMA (OLA)

Target Specific PCR







SNP typing, CF, CYP 450, RVP Gene expression, SNP typing, CNV, etc RVP FAST, GPP FAST

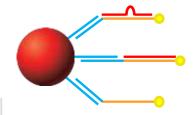
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miRNA Analysis:

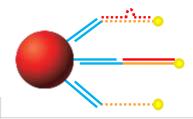
Nuclease Protection Approach



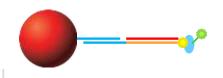
Step-down Probe Hybridization – DNA/RNA chimeric probes hybridize to target miRNAs during incremental reductions in annealing temperature. **2 hours**



Microsphere Hybridization – miRNA-chimeric probe complexes are hybridized to microspheres. **30 minutes**



RNase Digestion – Excess probes, single-stranded RNAs and mismatched probes are digested. Only perfectly-matched probes are protected.
30 minutes



SAPE Incubation – A brief incubation with streptavidin-conjugated
R-Phycoerythrin (SAPE) incorporates reporter molecules.
30 minutes



Detection – Targets of interest are quantified on a Luminex instrument.< 5 hours total to results</td>Luminex.



LMA Multiplex Gene Expression Assays

cDNA required for LMA Gene Expression Assay

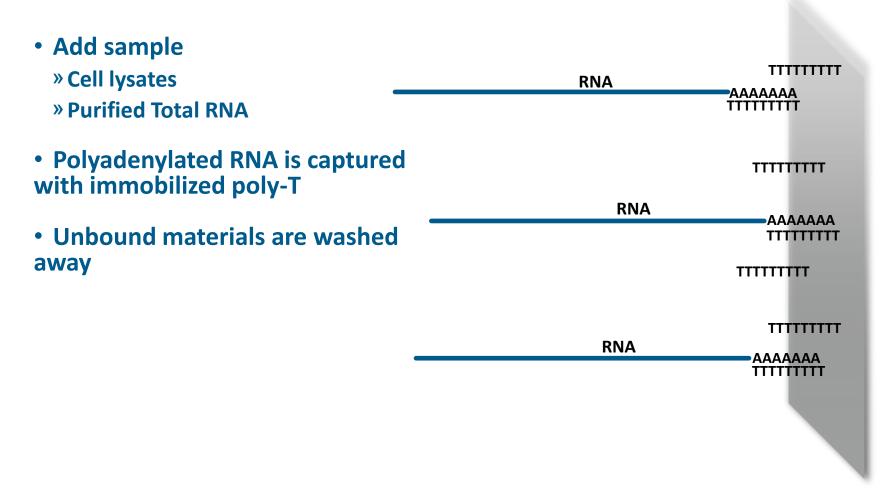
LMA = Ligation Mediated Amplification

• Oligo-dT primed cDNA synthesis.

✓ Can generate cDNA only from poly-A tailed mRNAs
 ✓ Can be optimized with poly-A capture methods
 ✓ Can be more sensitive than TaqMan and other PCR methods

O Random primed cDNA synthesis.
 ✓ Can generate cDNA from all RNA species
 ✓ Can also detect non-ploy-A transcripts
 ✓ Slightly shorter protocol than poly-A capture method

LMA Poly-A mRNA Capture Approach

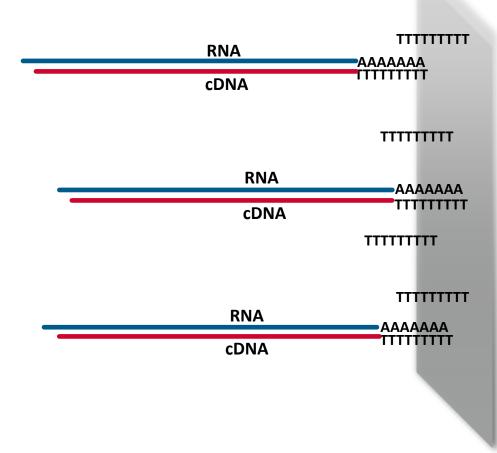


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LMA Poly-A mRNA Capture Approach

• RT Synthesis of 1st strand cDNA

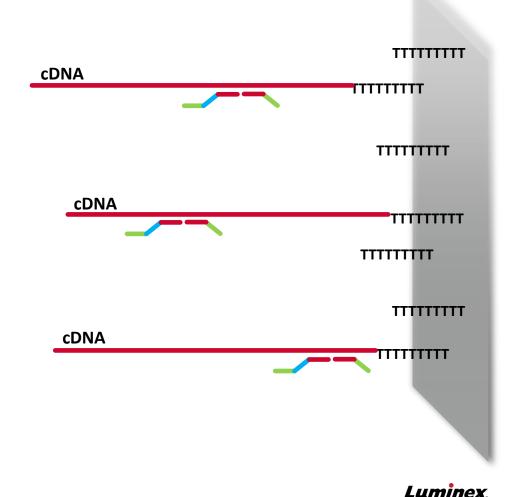


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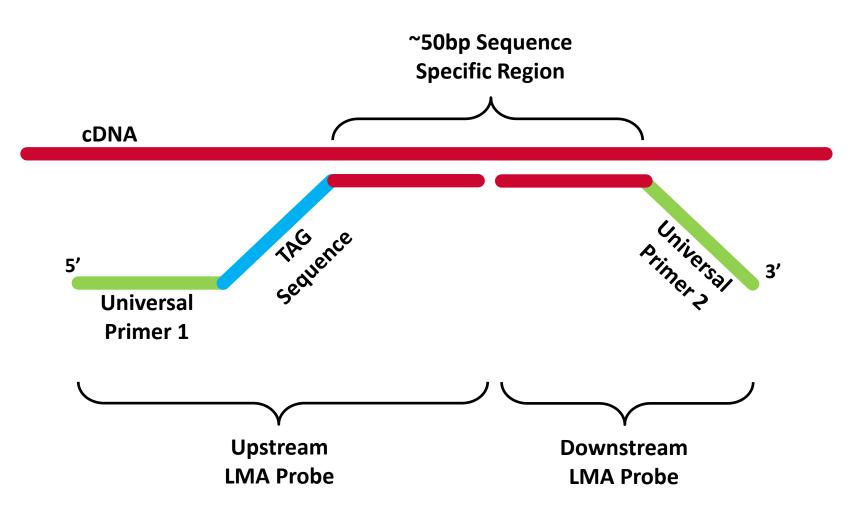
LMA Poly-A mRNA Capture Approach

Probe Hybridization

- Probes added
 - » 2 probes per transcript
 - » ~25 bases of target-specific sequence per probe
- Overnight or shorter **Hybridization**



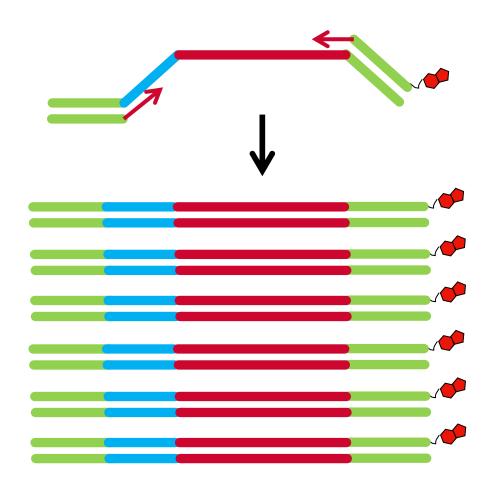
LMA Probe Design



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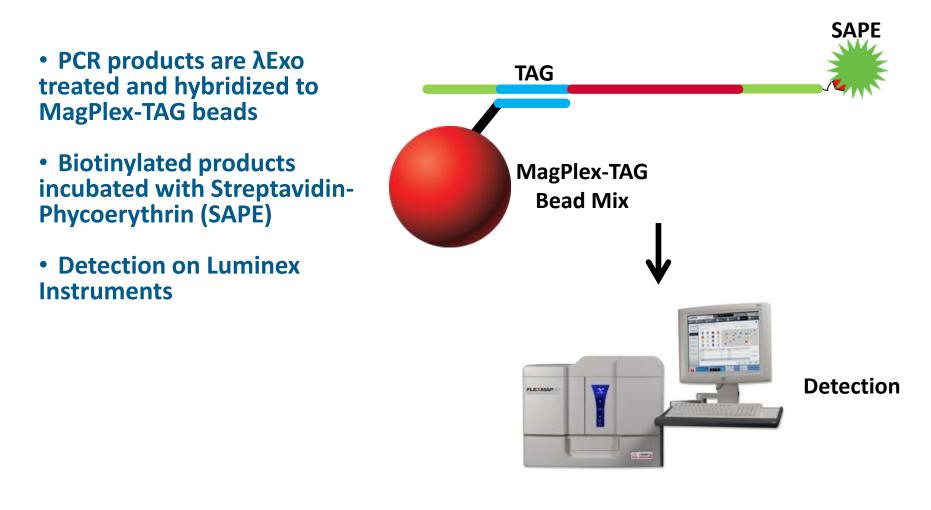
LMA Poly-A mRNA Capture Approach PCR Amplification

- Same universal primers used for all probe sets
- No size bias: all PCR products are same size
- Downstream primer is biotinylated

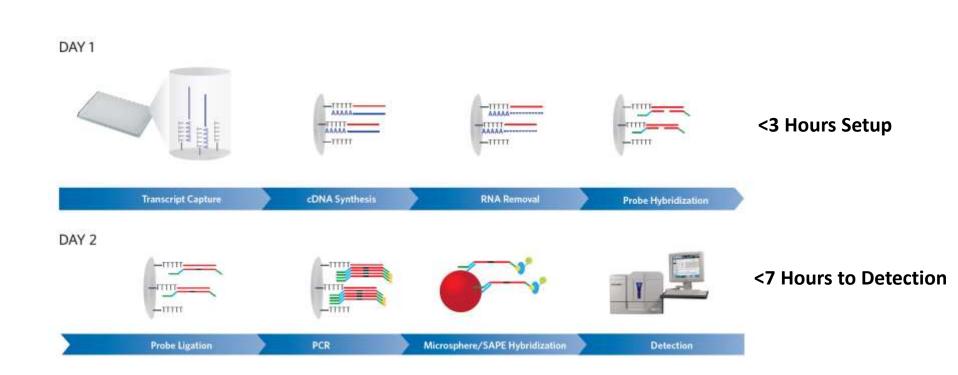


LMA Poly-A mRNA Capture Approach

Bead Hybridization and Detection



LMA Poly-A mRNA Capture Approach Workflow





Literature References

Peck et al. 2006; A method for high-throughput gene expression signature analysis.

Haining et al. 2008; High-throughput gene expression profiling of memory differentiation in primary human T cells.

Shao, X. J., et al. 2011; (Chinese) Development of a Beadbased Liquid Array for Analysis of Gene Expression Profiling.

Reijans, M., et al. 2008; RespiFinder: a new multi parameter test to differentially identify fifteen respiratory viruses.

Used for L1000 platform by GENOMETRY, INC



Luminex Partner Options for Genomic assays



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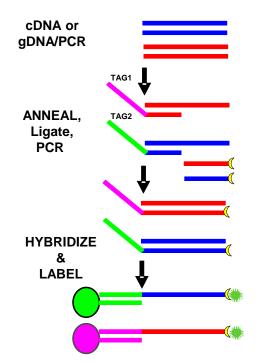


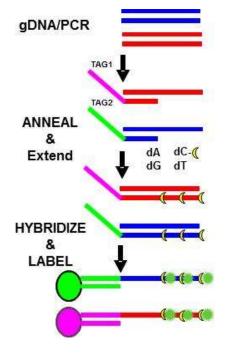
Luminex Based Multiplex SNP Genotyping Assays

Different SNP Chemistries

LMA (OLA)

Allele Specific Primer Extension





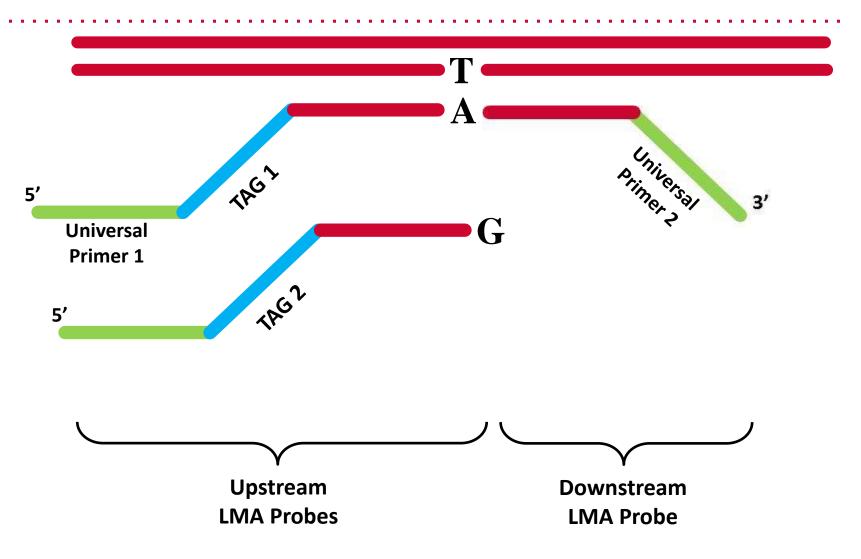




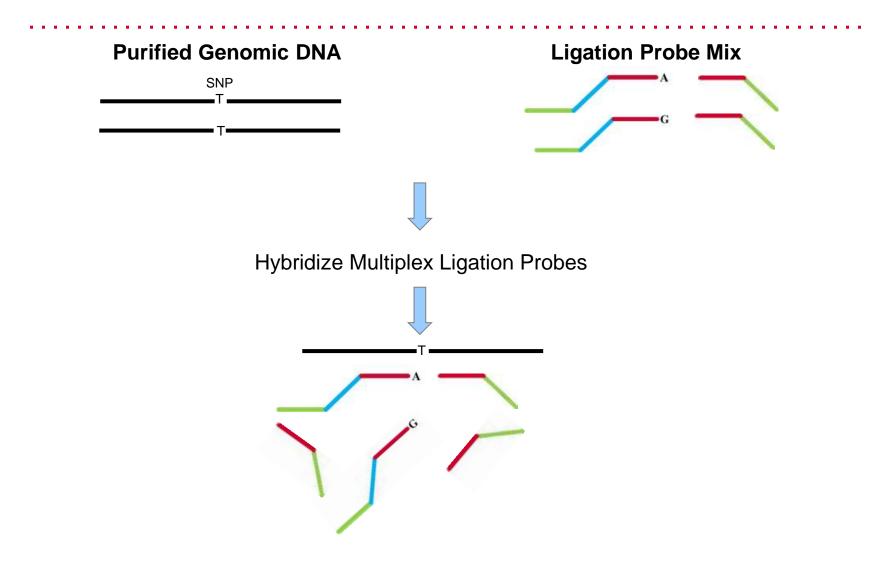
LMA Genotyping Assay

LMA Probe Design

aka... LMA, MLPA, MolPCR

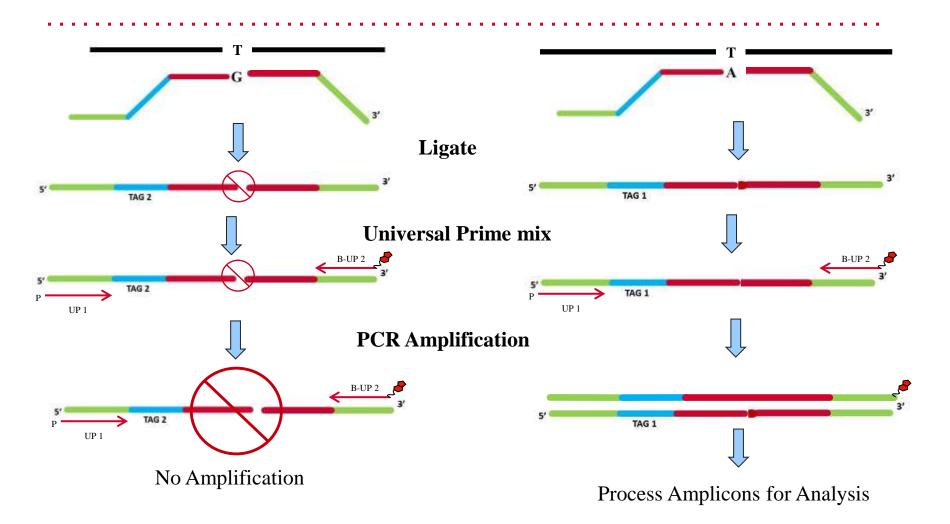


LMA Single Nucleotide Analysis

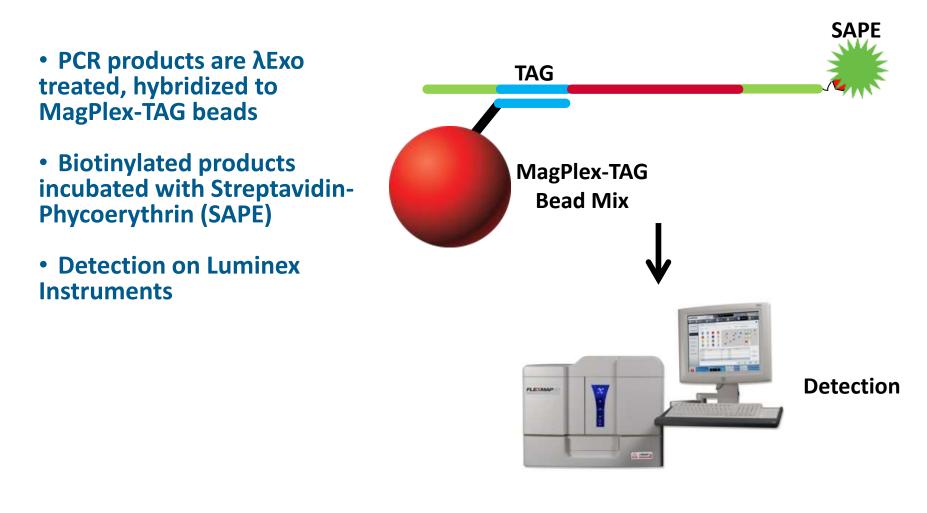




LMA Single Nucleotide Analysis



Bead Hybridization and Detection



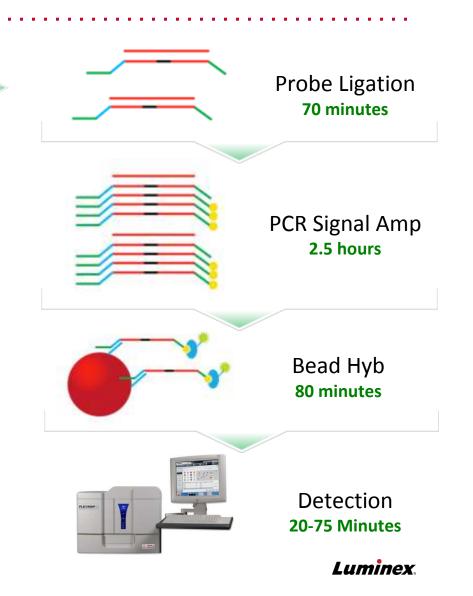
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LMA Genotyping Workflow



Probe Hybridization Overnight or less Incubation

- Overnight or shorter Incubation Followed by Data Collection
- Automation-Friendly Workflow
- PCR Uses Universal Primers for Unbiased Signal Amplification
- Optimized for Fast Read Times



Literature References

Kaderali et al 2003; Primer-design for multiplexed genotyping

Zhi, J. and E. Hatchwell 2008; Human MLPA Probe Design (H-MAPD): a probe design tool for bothelectrophoresis-based and bead-coupled human multiplexligation-dependent probe amplification assays

Milosevic et al. 2010 Development and validation of a comprehensive mutation and deletion detection assay for SDHB, SDHC, and SDHD

Deshpande et al. 2010; A rapid multiplex assay for nucleic acid-based diagnostics

Song et al. 2010; Simultaneous Pathogen Detection and Antibiotic Resistance Characterization Using SNP-Based Multiplexed Oligonucleotide Ligation-PCR (MOL-PCR)

Stucki et al. 2012; Two new rapid SNP-typing methods for classifying *Mycobacterium tuberculosis* complex into the main phylogenetic lineages

Deshpande & White 2012; Multiplexed nucleic acid-based assays for molecular diagnostics of human disease



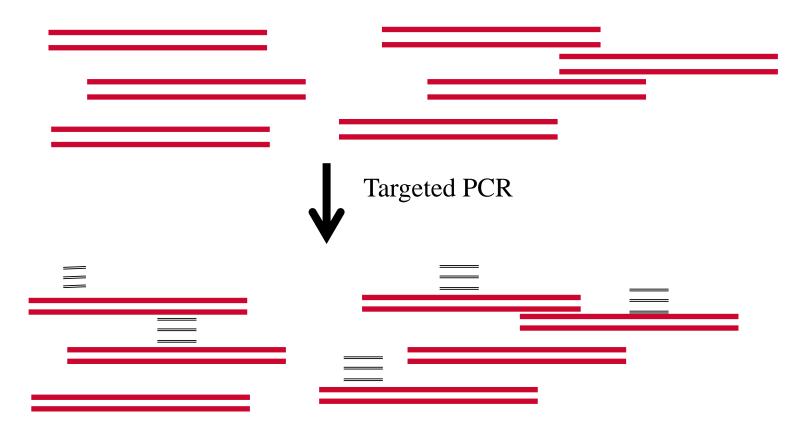


OLA (LDR-FMA) Genotyping Assay

OLA (LDR-FMA) Genotyping

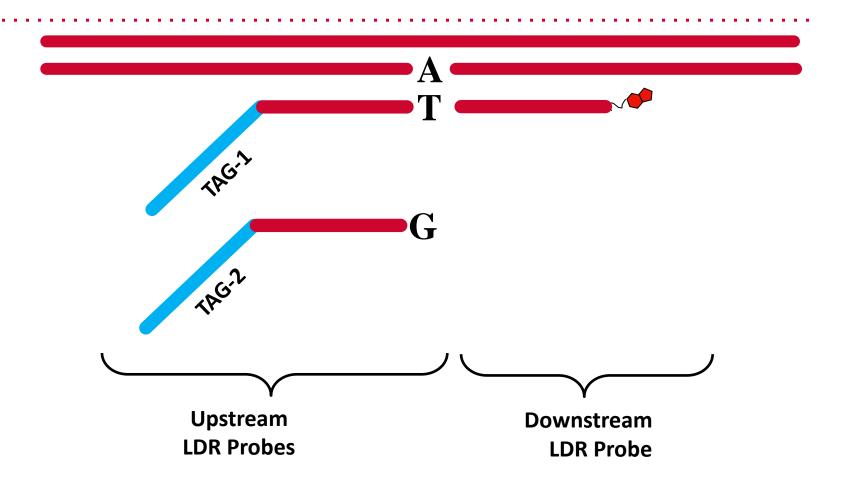
OLA = Oligo Ligation Amplification LDR-FMA = ligase detection reaction - fluorescent microsphere assay

Generate ligation probe target regions of interest with PCR

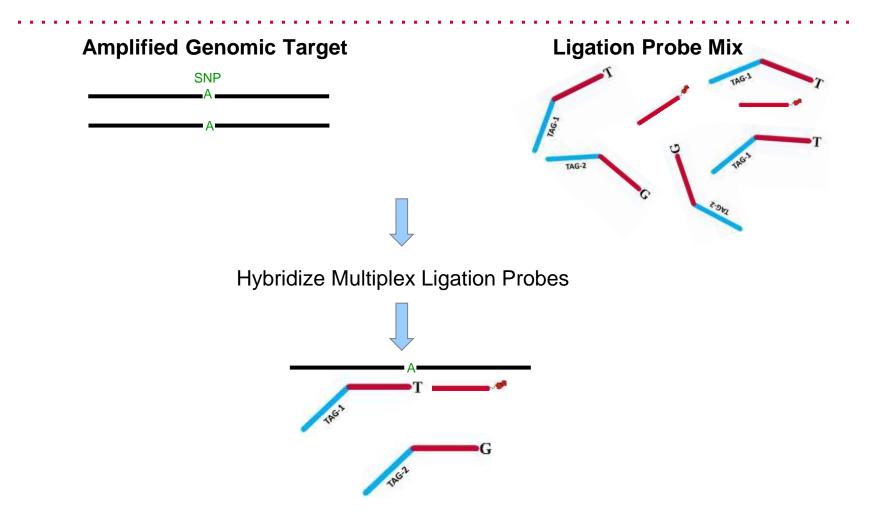




OLA (LDR-FMA) Probe Design

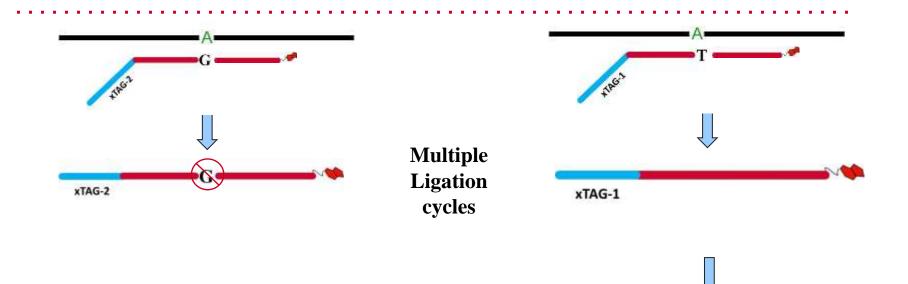


OLA Single Nucleotide Analysis





OLA Single Nucleotide Analysis

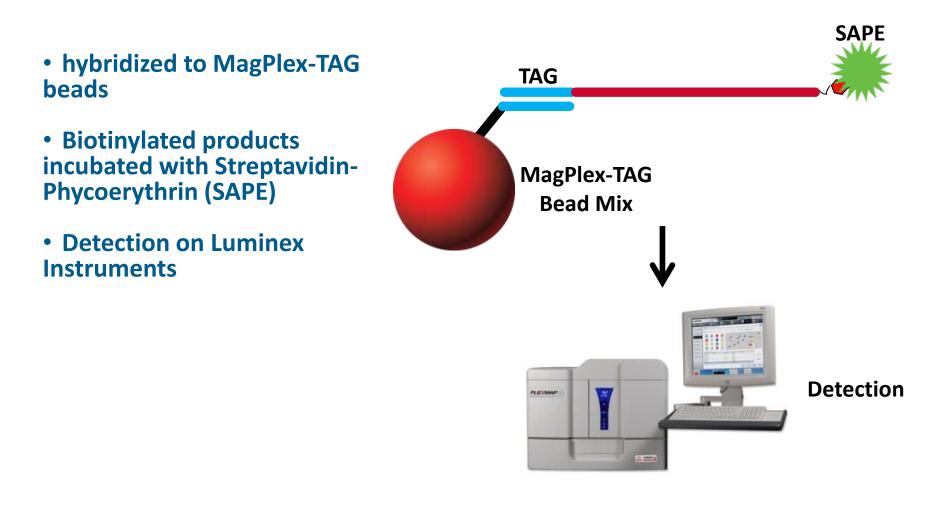


Process Ligated Probes for Analysis

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Bead Hybridization and Detection



Literature References

Carnevale et al. 2007; A Multiplex Ligase Detection Reaction-Fluorescent Microsphere Assay for Simultaneous Detection of Single Nucleotide Polymorphisms Associated with *Plasmodium falciparum* Drug Resistance

Bruse et al. 2008; Improvements to bead-based oligonucleotide ligation SNP genotyping assays

Mehlotra et al. 2011; Chemokine (C-C motif) receptor 5 -2459 genotype in patients receiving highly active antiretroviral therapy race-specific influence on virologic success

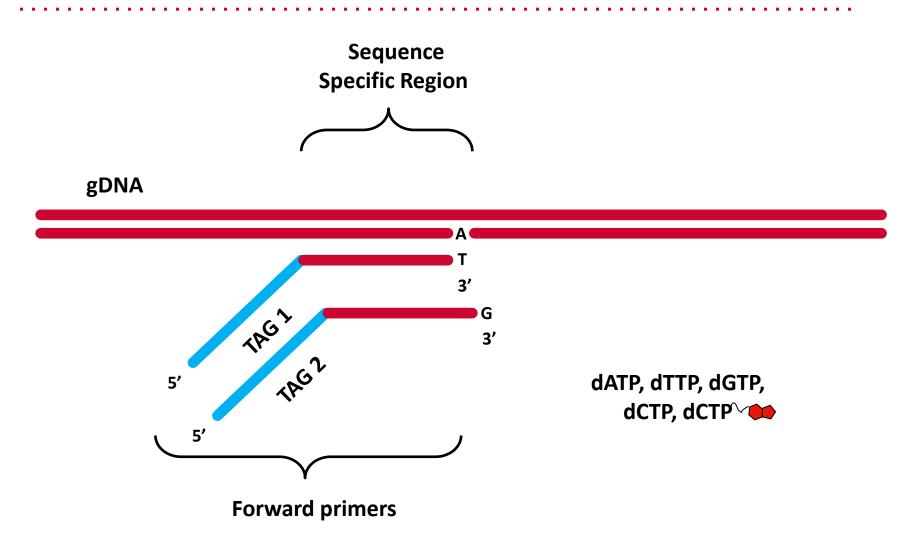
Henry-Halldin et al. 2012; Multiplex Assay for Species Identification and Monitoring of Insecticide Resistance in *Anopheles punctulatus* Group Populations of Papua New Guinea



ASPE SNP Typing Assays

ASPE Probe Design

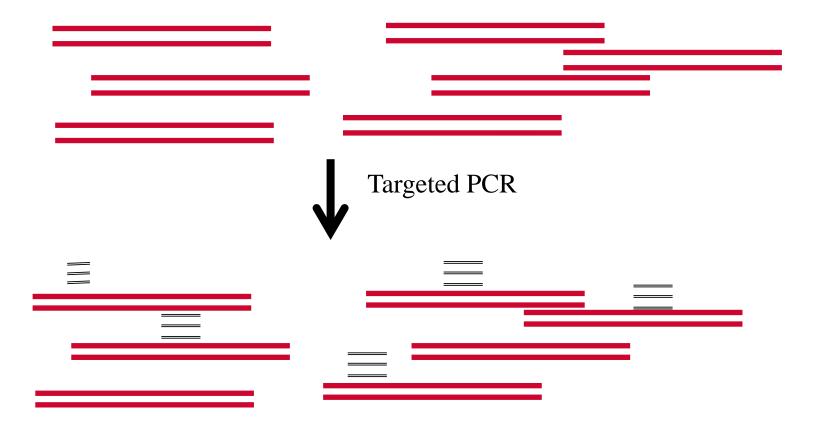
With one Biotinylated-Nucleotide Chemistry



ASPE SNP Genotyping

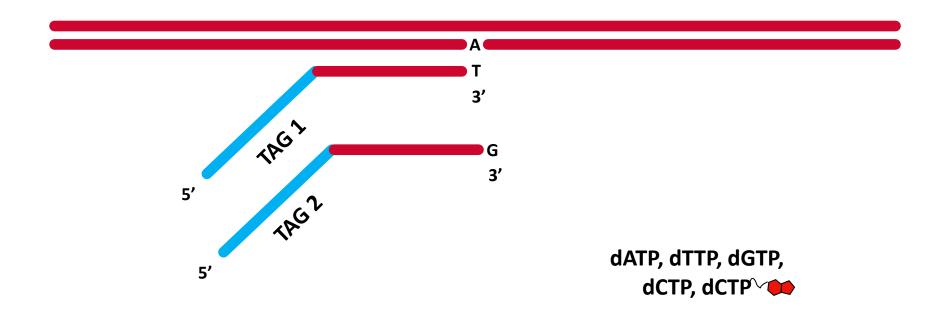
gDNA Target Amplification

Amplify probe target regions of interest with PCR

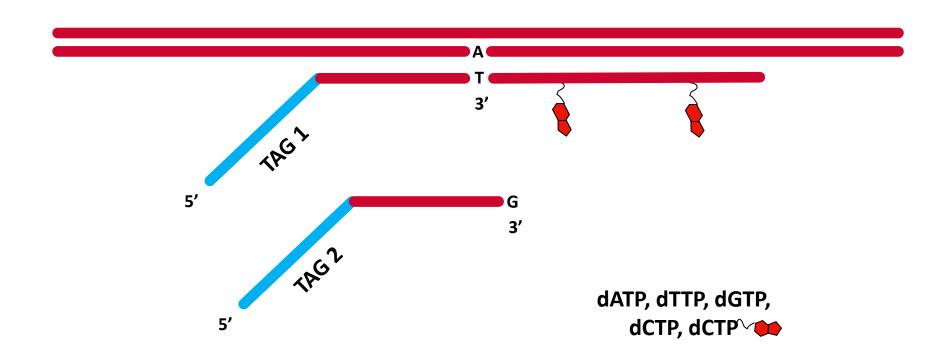




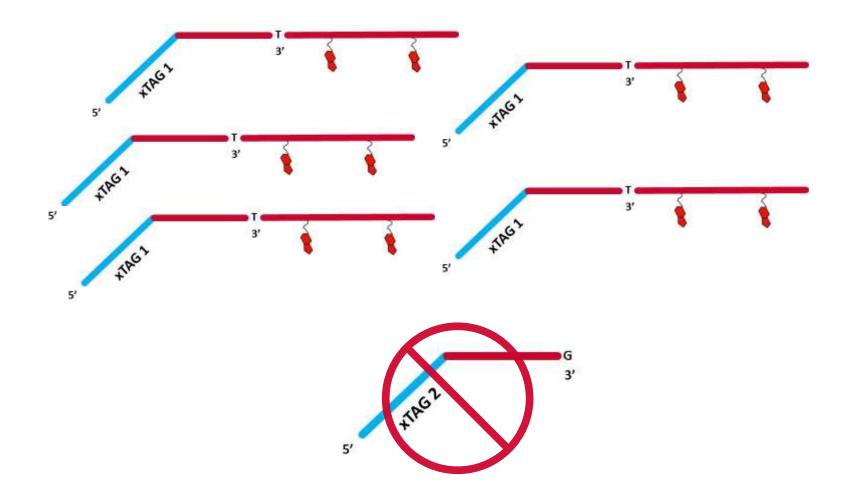
Primer Extension on amplified regions



Primer Extension on amplified regions

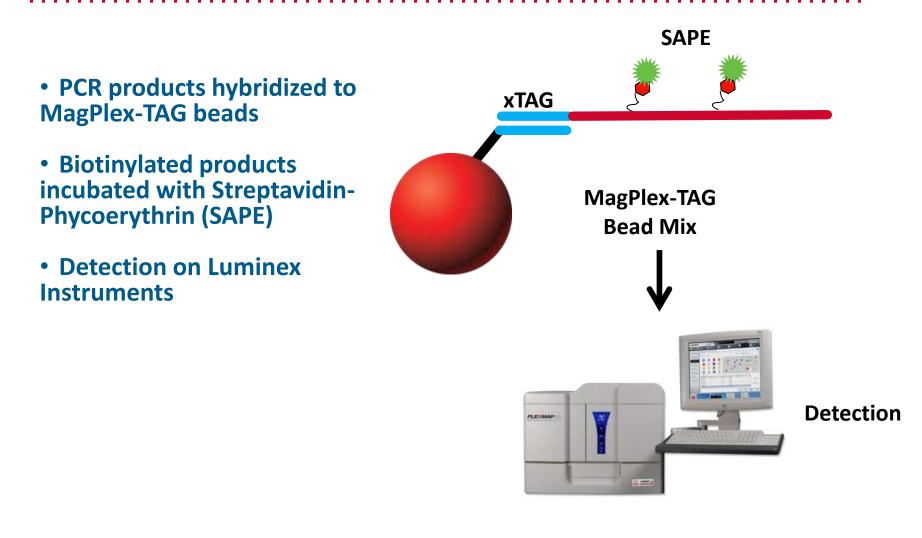


Primer with complimentary base is extended and biotynalated



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With Biotinylated-nucleotide.



Literature References

Strom et al. 2005; Technical validation of a multiplex platform to detect thirty mutations in eight genetic diseases prevalent in individuals of Ashkenazi Jewish descent

Koo et al. 2007; Multiplexed genotyping of ABC transporter polymorphisms with the Bioplex suspension array

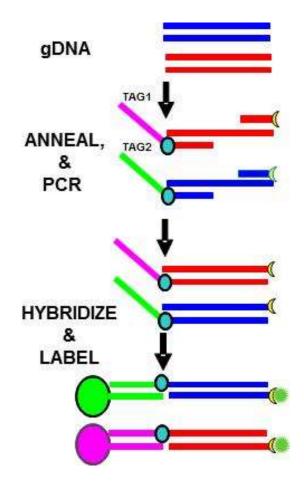
Li et al. 2011; A novel liquidchip platform for simultaneous detection of 70 alleles of DNA somatic mutations on EGFR, KRAS, BRAF and PIK3CA from formalin-fixed and paraffinembedded slides containing tumor tissue





Target Specific PCR Chemistry

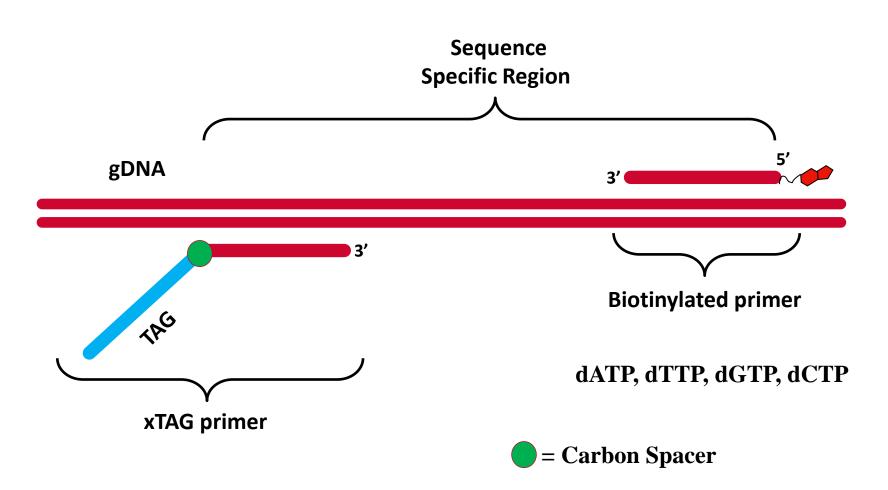
Target Specific PCR TS-PCR Chemistry





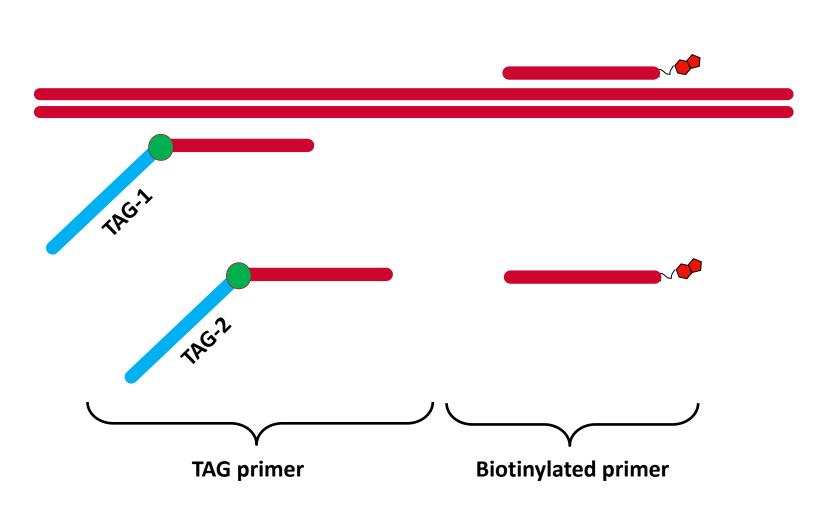
TS PCR Probe Design

With Biotinylated primer and standard nucleotides

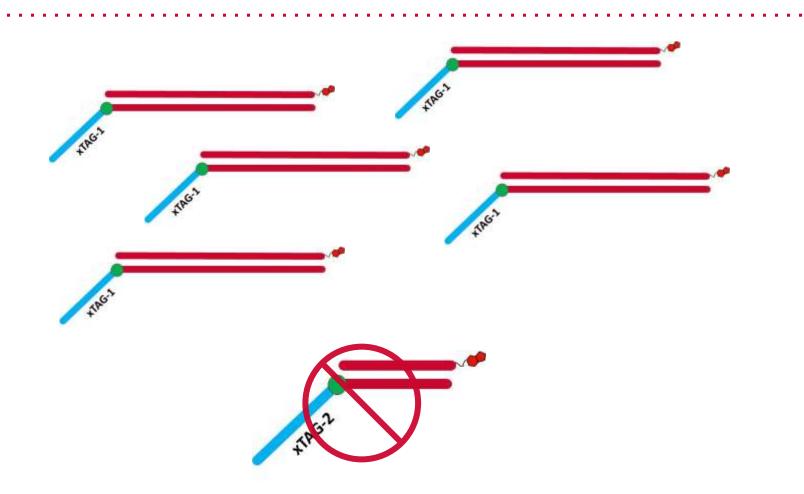


TS PCR Probe Design

With Multiplex Probe mix

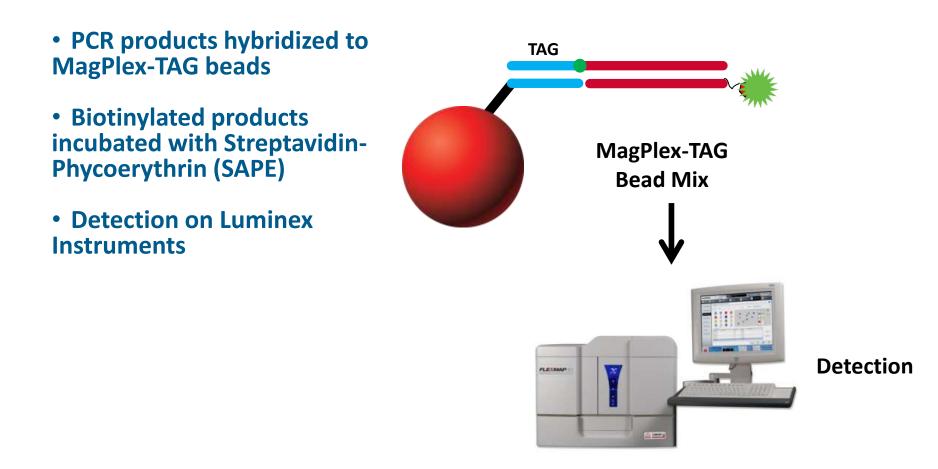


Multiplex Amplification Products





Sequence Detection



FAST PCR Assay Characteristics

Summary

Easy and Fast

- Simple sample processing from a number of different sample types such as swabs, stool, cultures, etc.
- Results in a few hours.

• Used in Diagnostics for;

- <u>Detection of 8 viruses and subtypes</u> of Influenza A, Influenza A subtype H1, Influenza A subtype H3, Influenza B, Respiratory Syncytial Virus (RSV), Human Metapneumovirus (hMPV), Rhinovirus, and Adenovirus.
- Detection of 11 gastrointestinal pathogens and subtypes of bacteria, viruses, and parasites of *C. difficile*, norovirus, *E. coli*, *Salmonella*, rotavirus A, *Campylobacter*, and *Shigella*.



Multiplex microRNA Assay: Nuclease protection Approach

Multiplex miRNA Nuclease protection Assay Characteristics

• Extremely Specific Results:

»Can discriminate single base mismatches anywhere in mature miRNA sequences

• Fast Results:

»From total RNA to results in less than 5 hours with less than30 minutes of handling

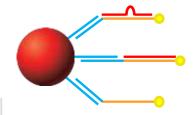
• Unbiased Sensitivity:

»No amplification or sample labeling required, yet data from 50-500ng total RNA with single nucleotide difference resolution

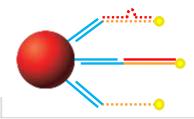
Workflow



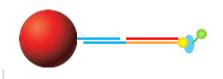
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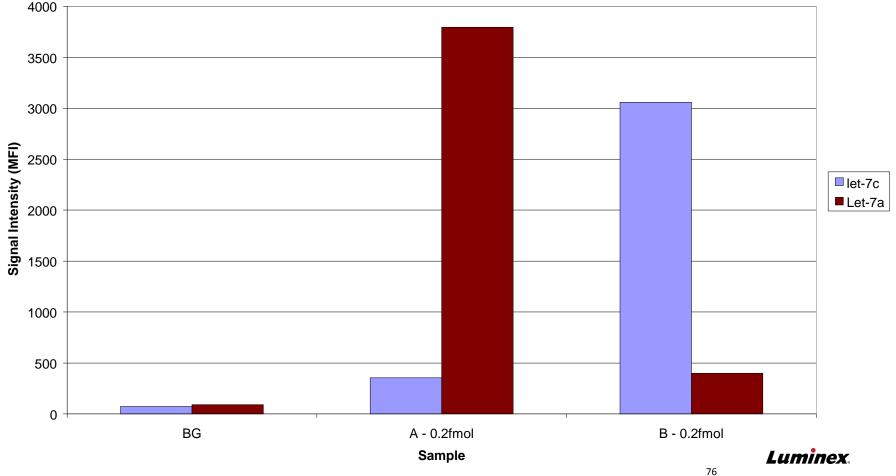
Detection – Targets of interest are quantified on a Luminex instrument.< 5 hours total to results</td>Luminex.

Specificity

Single Base Mismatch Discrimination

hsa-let-7a UGAGGUAGUAGGUUGUAUAGUU hsa-let-7c UGAGGUAGUAGGUUGUAUGGUU

Discrimination of single base mismatch near ends!

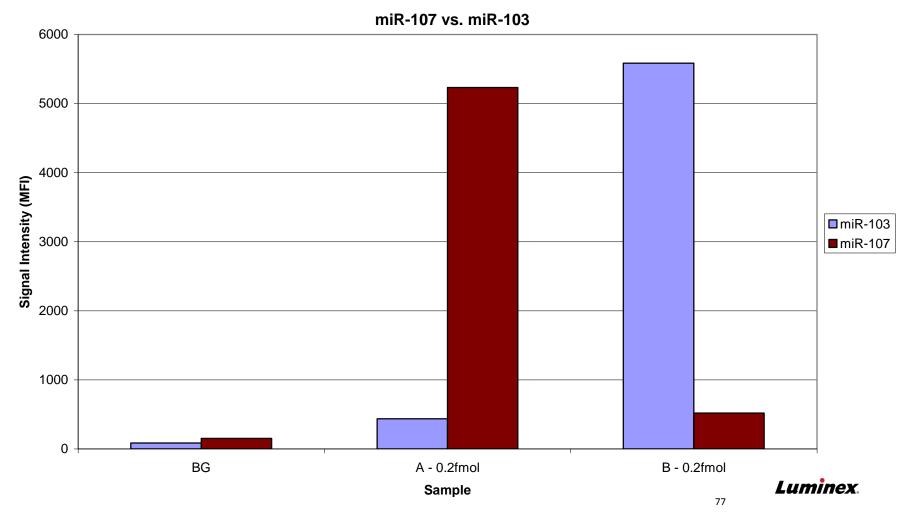


Specificity

Single Base Mismatch Discrimination

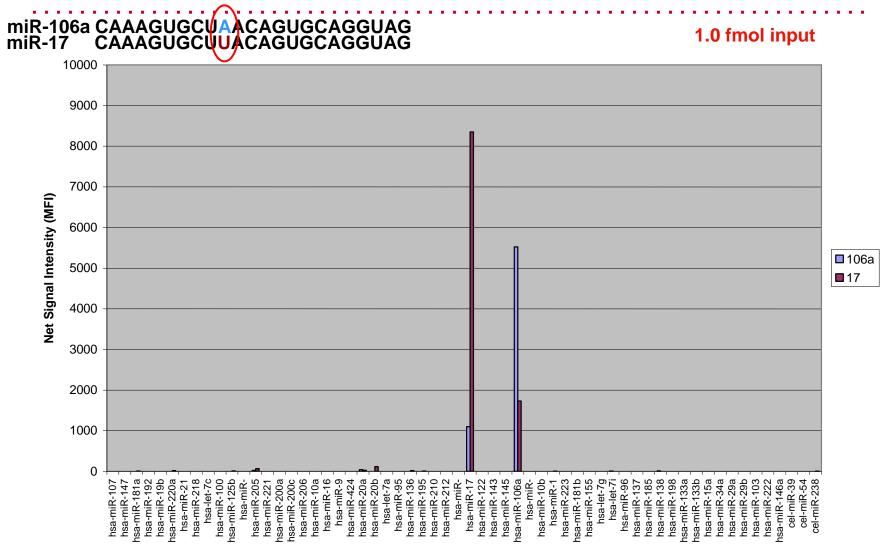
hsa-miR-107 AGCAGCAUUGUACAGGGCUAUCA hsa-miR-103 AGCAGCAUUGUACAGGGCUAUGA

Discrimination of single base mismatch near ends!



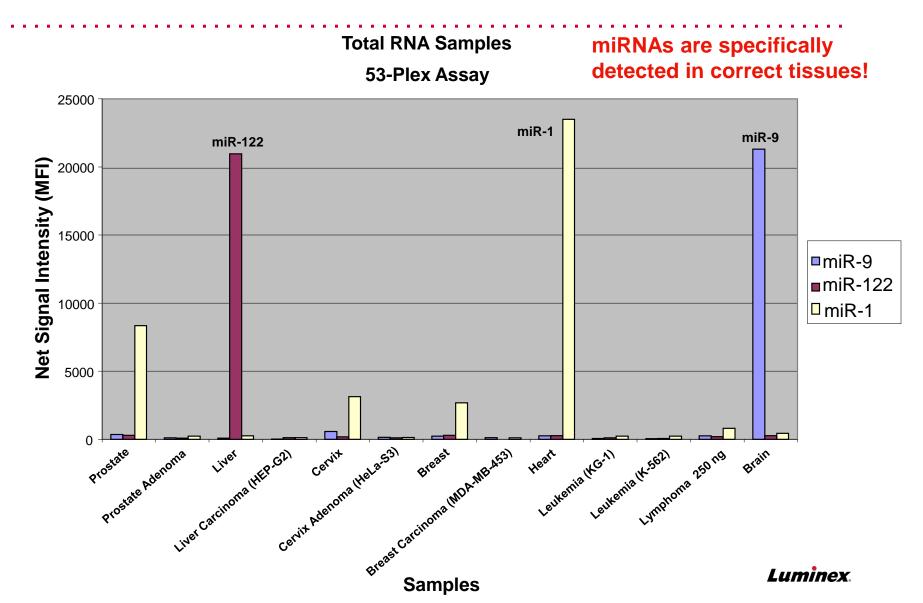
Specificity for Targets in Complex mix

Closely-Related miRNAs in 60 Plex

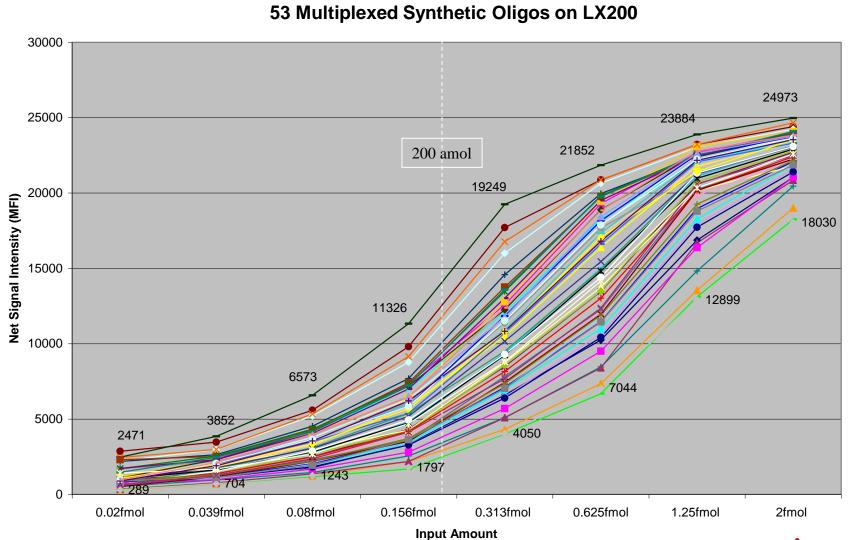


Specificity in Tissue Samples

Correct Detection of Tissue-Specific miRNAs

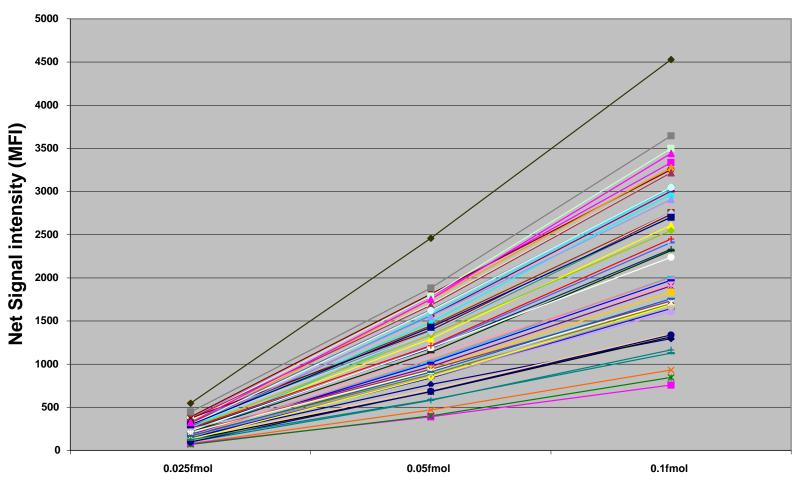


Sensitivity in Complex mix 200 amol Molecules or Less – Unamplified!



Sensitivity limits

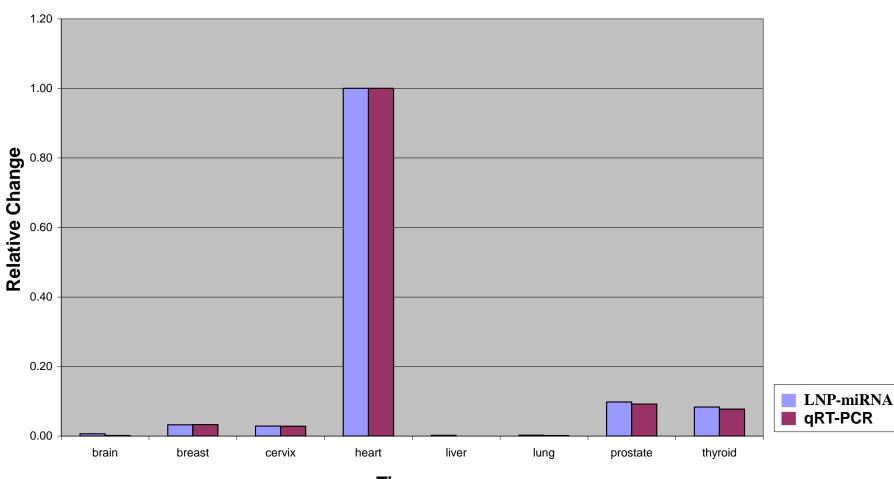
Titration below 100 amols – Unamplified! 53 plex Synthetic Oligos on FlexMAP 3D



Input Amount (fmol) (Note Expanded Scale)



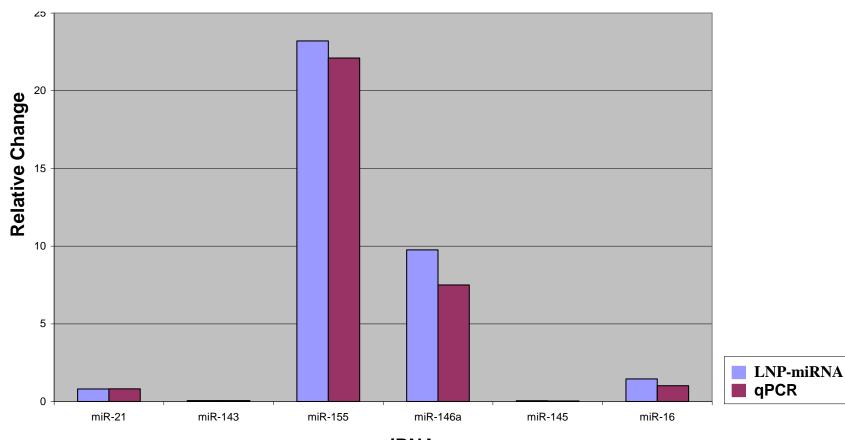
Accuracy vs. qPCR Correlation to qRT-PCR for miR-133a



Tissue

Accuracy vs. qPCR

Correlation to qPCR of 6 microRNAs Fold Change of Tumor vs. NAT - Lymphoma Samples



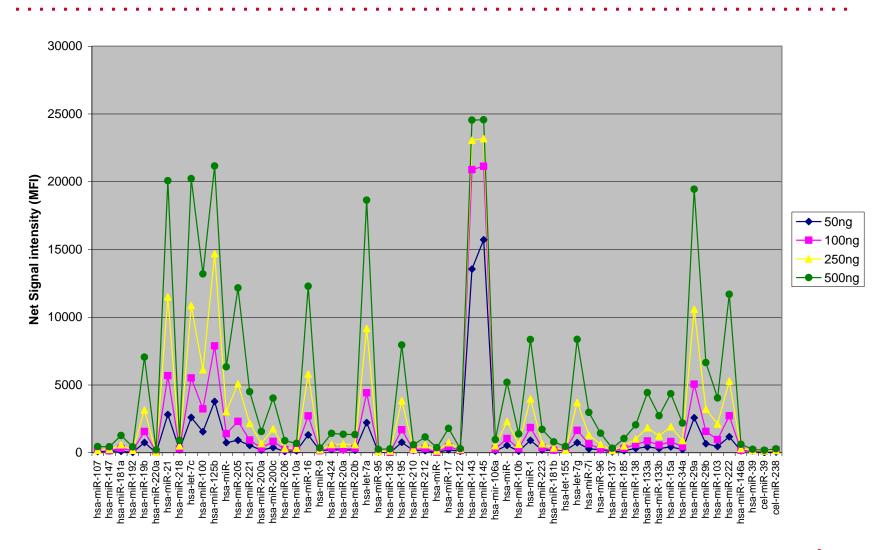
miRNA

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Accuracy at Different RNA Inputs

Consistent Results with Varying Total Prostate RNA



miRNA Assay Characteristics

Summary

• Fast, Easy and Unbiased

»Only 30 Minutes of Handling
 »No Sample Labeling or Amplification
 »Time to Results <5 hours

Solid Performance

»Reproducible: CVs <15%

»Sensitive: 50-500ng Total RNA Input

»Highly Specific: Discriminates Single Base Mismatches

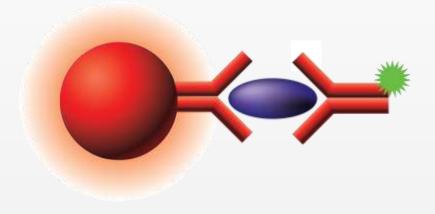
• Flexible

»Multiplexing of >80 miRNAs/well possible

»Design your own custom probes



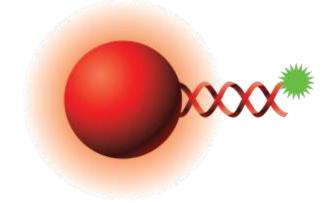
Proteomic Assays



xMAP Applications

An Incredibly Flexible Platform

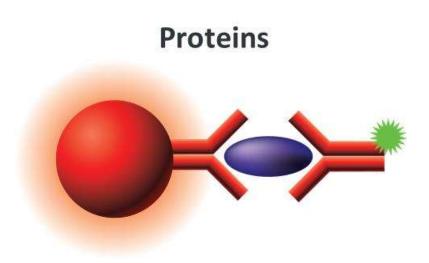
Nucleic Acids



- Gene ExpressionmicroRNA Profiling
- Genotyping »SNP

»CNV

»Sequence Detection



- Immunoassays » Sandwich Capture » Multiplex ELISA
- Receptor-Ligand/Protein interaction Assays
- Enzyme Substrate

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Multiplex Serological (ELISA) Assays

Multiplex ELISA

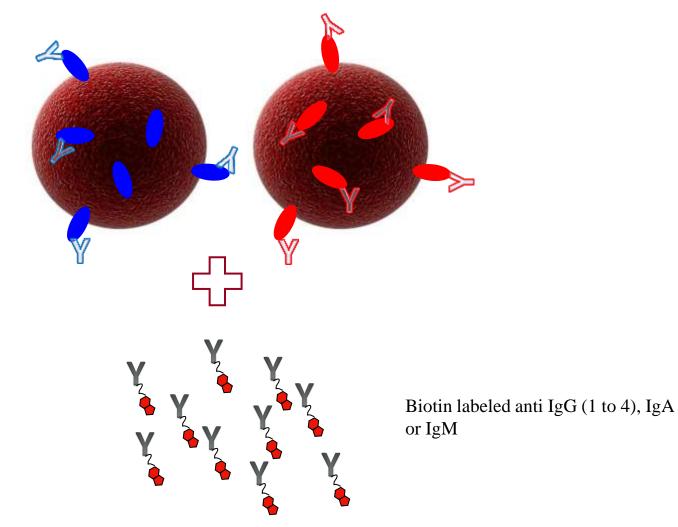
Mix beads with sera



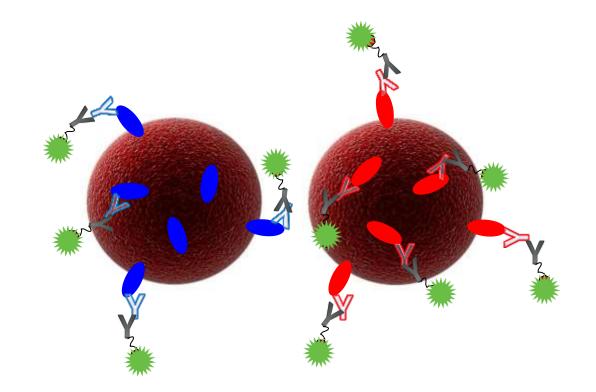


Multiplex ELISA

Mix Beads with Biotinylated Detection Antibody mix

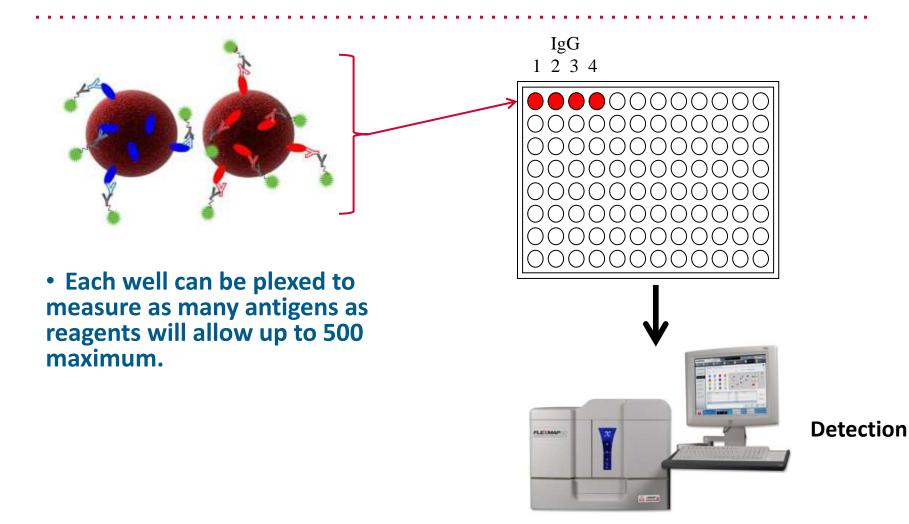


Multiplex ELISA Mix Beads with SA-PE



Can now determine if antibody response to antigen is IgG1, 2, 3, 4, or IgA or IgM

Multiplex Detection



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Literature References

Anderson, S., P. Wakeley, G. Wibberley, K. Webster and J. Sawyer (2011). "Development and evaluation of a Luminex multiplex serology assay to detect antibodies to bovine herpes virus 1, parainfluenza 3 virus, bovine viral diarrhoea virus, and bovine respiratory syncytial virus, with comparison to existing ELISA detection methods." Journal Of Immunological Methods 366: 79-88.

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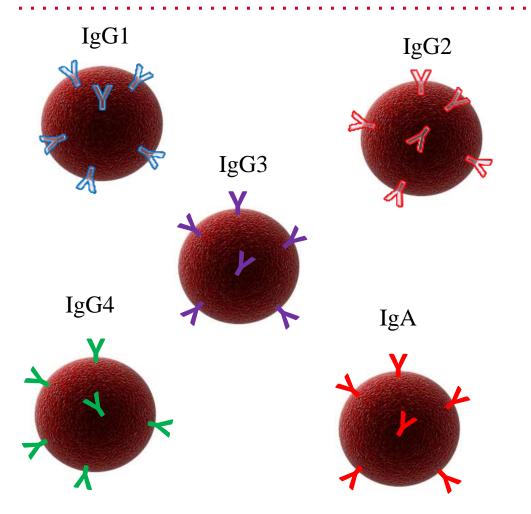
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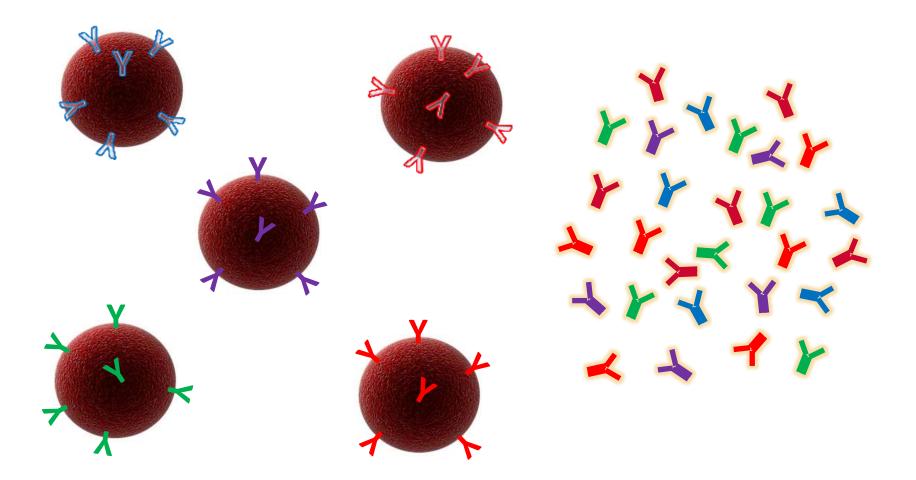


Multiplex Antigen Specific Ig ELISA assay Beads Coated with Capture Antibodies for Different Ig types



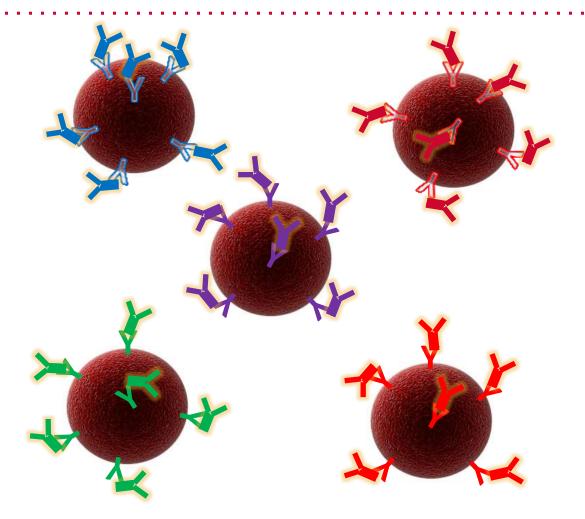


Beads with Capture Antibodies incubated with serum sample



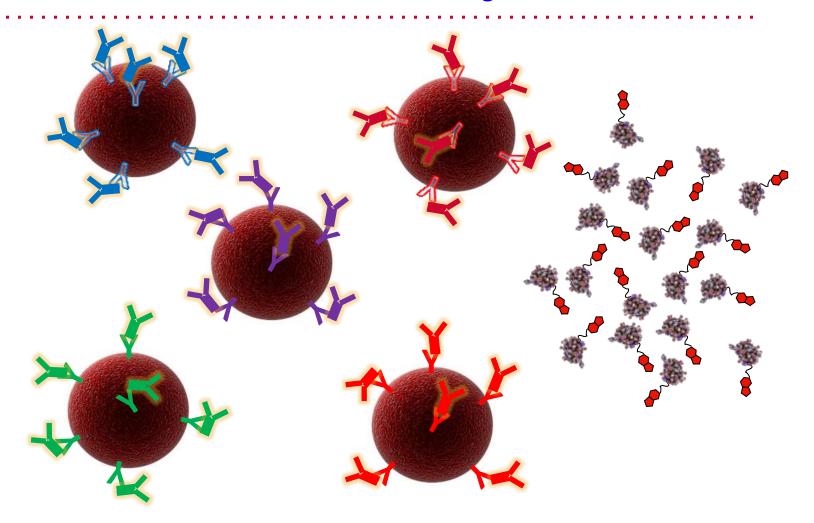


Beads Capture Antibodies of Different Ig types





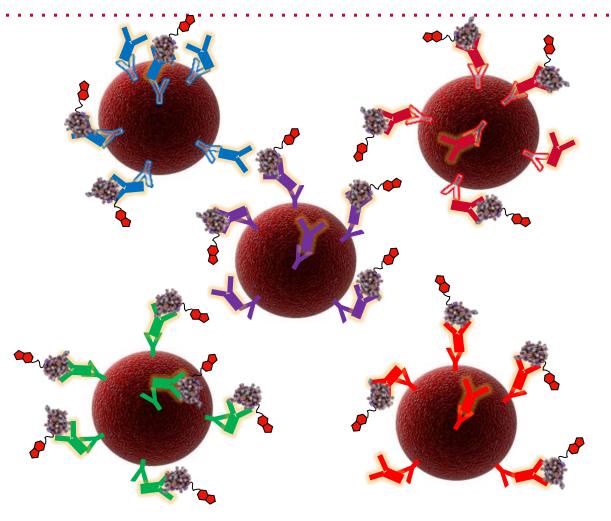
Beads Incubated with Biotin Labeled Antigen





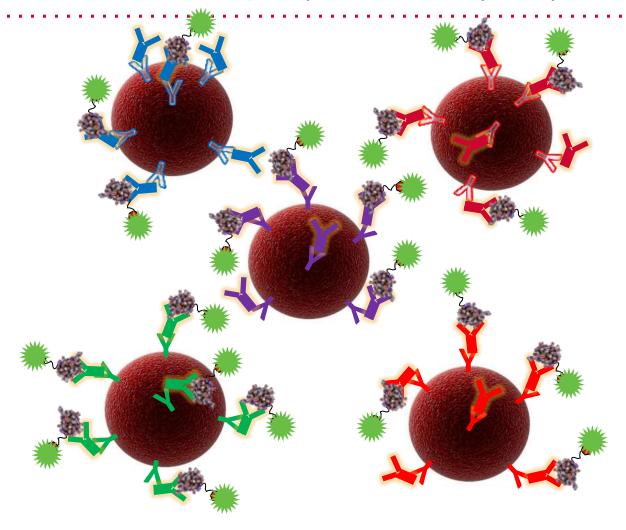


Different Ig types will bind Antigen



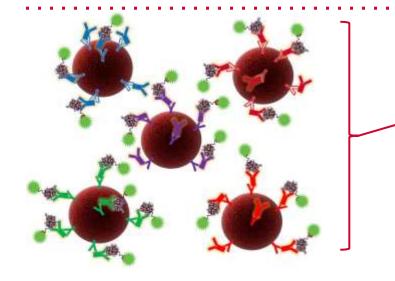


Incubate with SAPE (Streptavidin-R-Phycoerythrin)

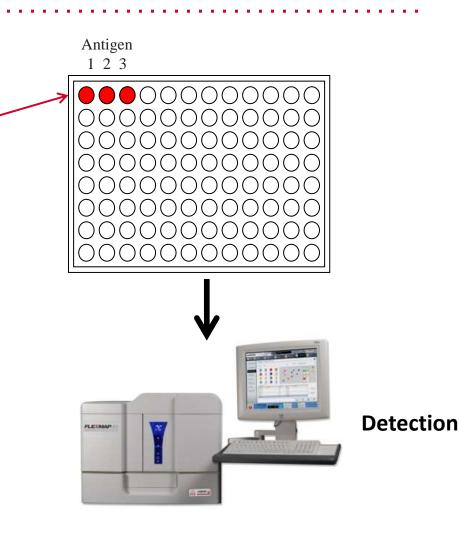




Multiplex Antigen Specific Ig ELISA assay Detection

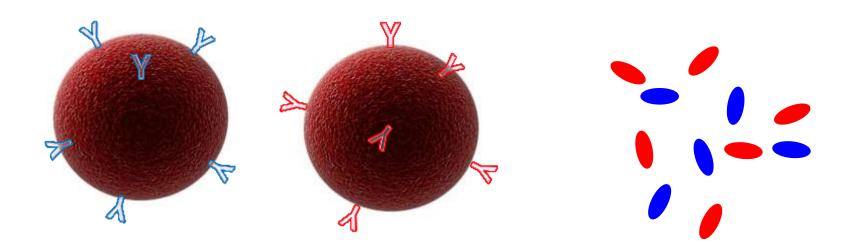


- Each well can be plexed to measure as many antigen specific Ig subtypes as needed
- Allows more rapid analysis of Ig type responses to multiple antigens in large numbers of samples.



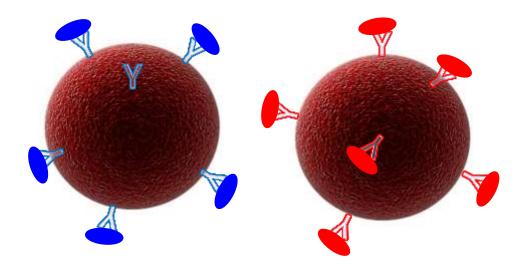


Beads Coated with Capture Antibodies Mixed with Sample



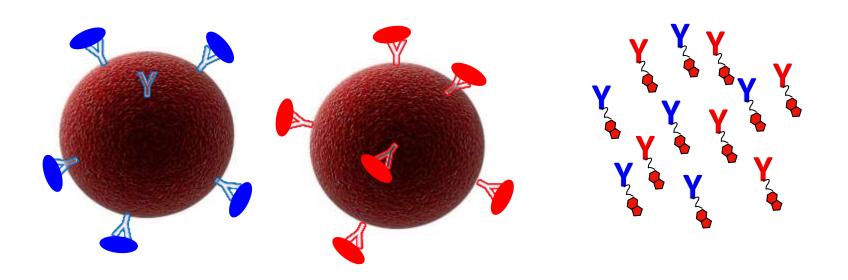


Targets Bind to Capture Antibodies



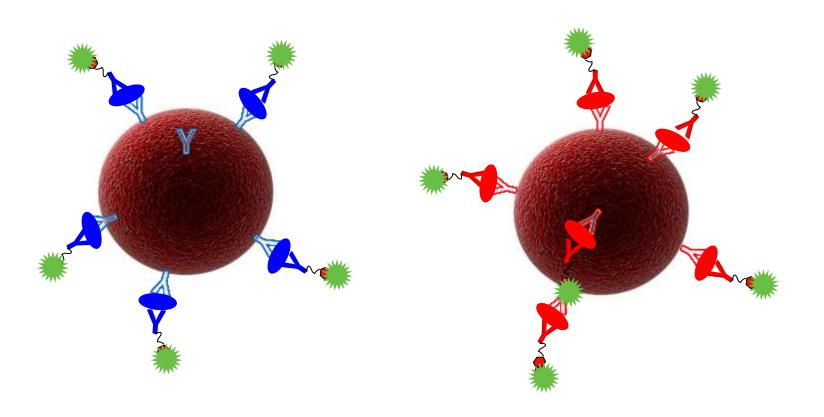


Mix with Labeled Target Specific Antibodies



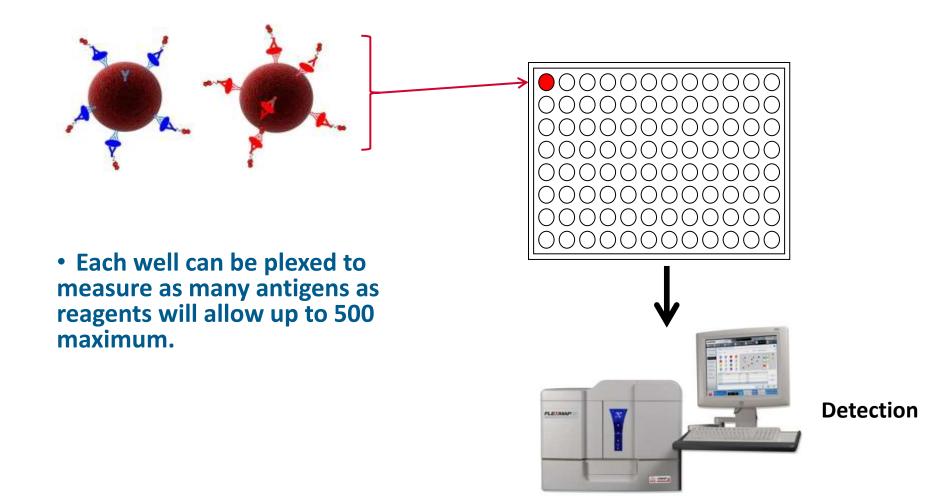


Mix with Labeled Target Specific Antibodies





Multiplex Detection



Literature References

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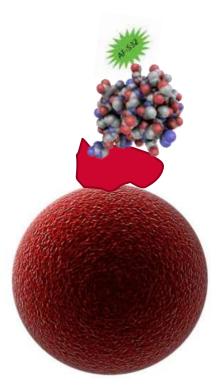


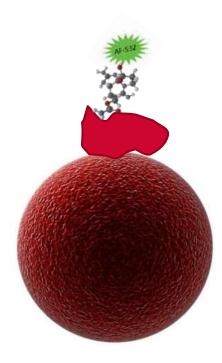


Protein Interaction and Receptor Ligand/Subatrate Assays

Types of Receptor Ligand and Protein Interaction Assays

Types of Receptor Ligand Interaction assays

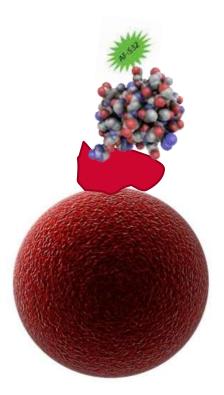


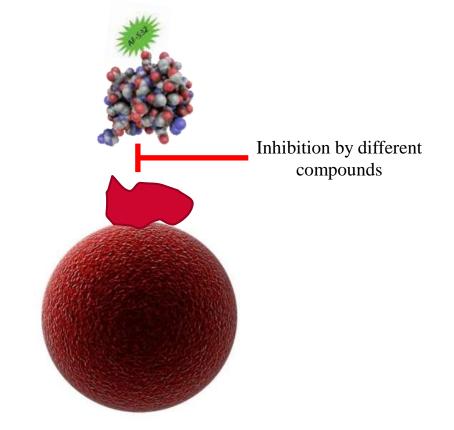




Types of Receptor Ligand and Protein Interaction Assays

Types of Receptor Ligand Interaction assays

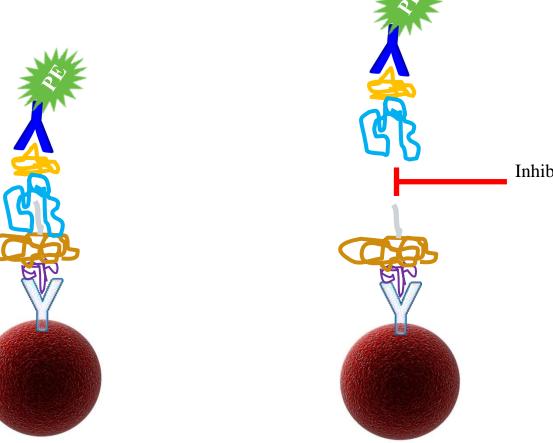






Types of Receptor Ligand and Protein Interaction Assays

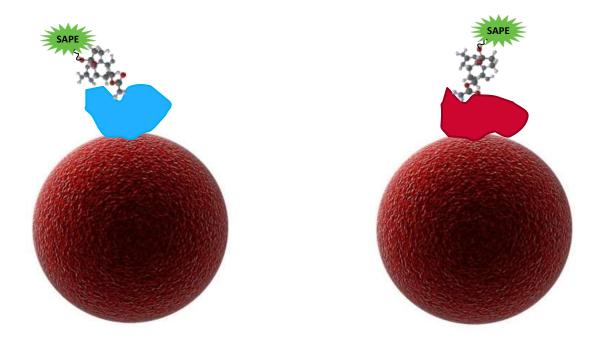
Types of Protein Interaction Assays



Inhibition by different compounds



A two plex assay: Labeled Substrate





A two plex assay: Inhibitor Treatment



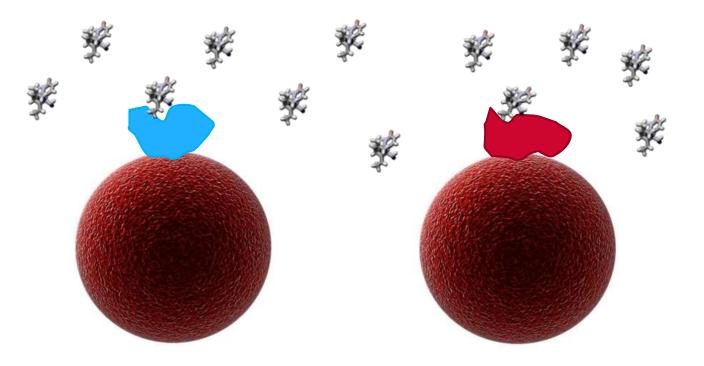
= inhibiting compound





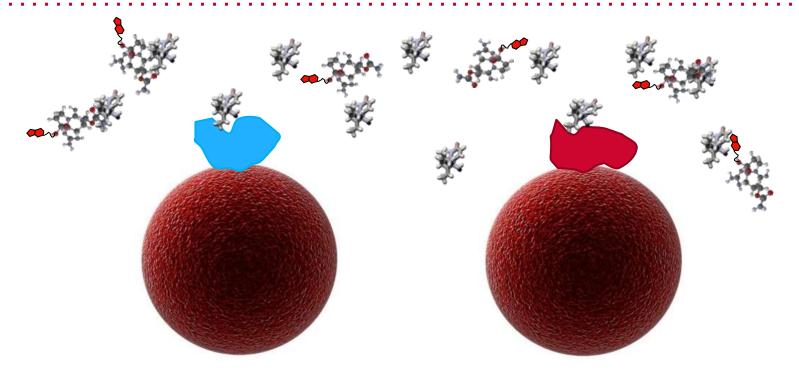


Inhibitor Treatment at Different Concentrations



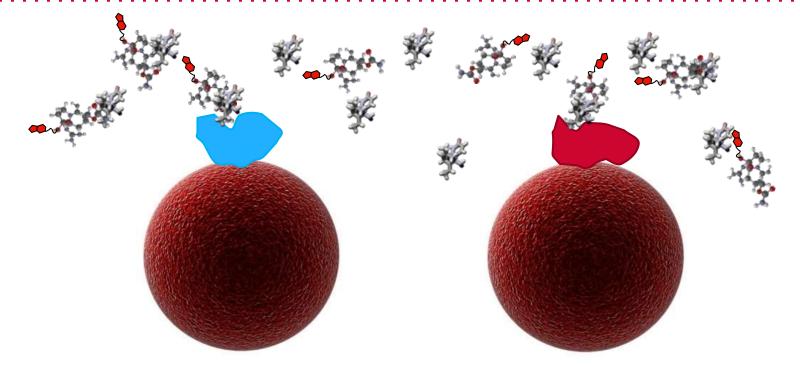


Add Labeled Substrate



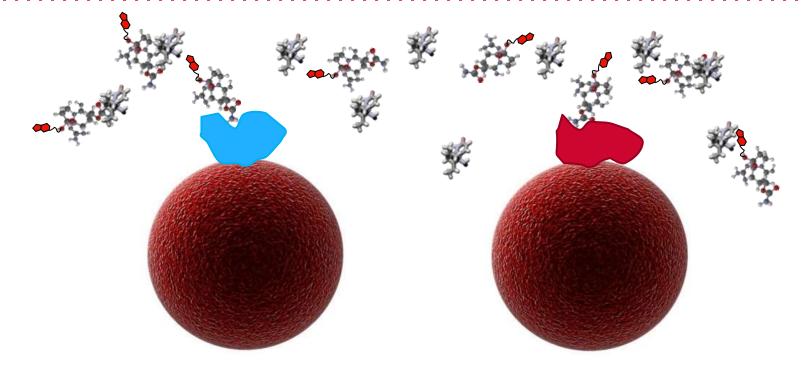


Labeled Substrate competes with Inhibitor



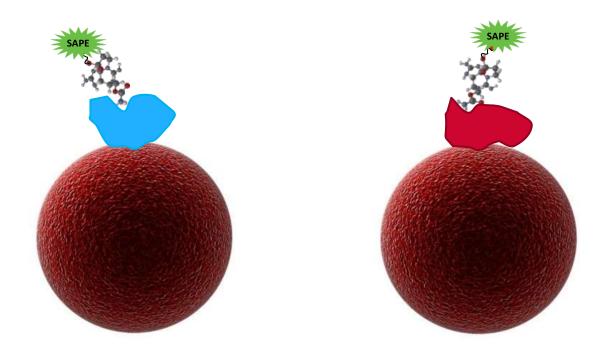


Wash out Excess Inhibitor and Labeled Substrate



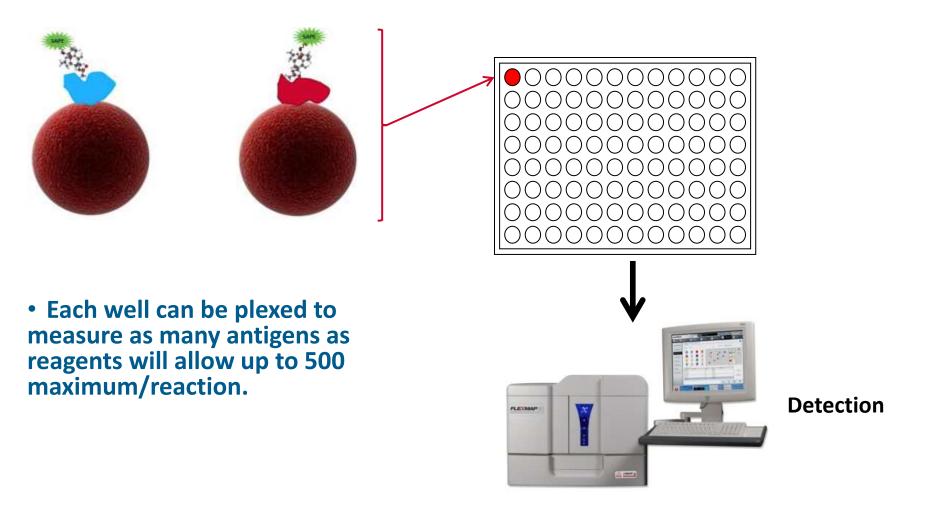


Hybridize with SAPE





Multiplex Detection



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Resources for Developing Multiplex Assays

Resources for Developing Multiplex Assays

1. The <u>Kit Finder</u>. Allows users to search for existing kits made by Luminex partners.

2. Papers with protocols of interest. Try the Luminex publication search tool.

3. The <u>xMAP Cookbook</u>. Helps users develop their own assays with all of their own reagents.



Resources for Developing Multiplex Assays

Luminex. XMAP* Technology

1st Edition

xMAP[®] Cookbook

A collection of methods and protocols for developing multiplex assays with xMAP Technology.



The xMAP Cookbook is a collection of easy to follow protocols with lists of reagents and equipment needed to develop you own Luminex based assay.





Interested in Getting Started?

For Instrument/Reagent Pricing Josh Jenkins Business Manager South East USA <u>jjenkins@luminexcorp.com</u> Cell: 512.202.9372

> For Technical Information Stephen Angeloni, PhD Sr. Field Application Scientist sangeloni@luminexcorp.com Cell: 512.348.1535

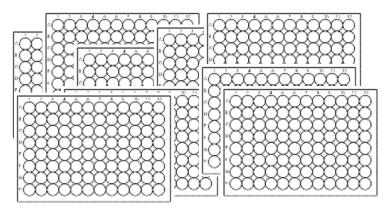
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Thank you for Attending

I will now take questions about converting your...

Single Plex Assays



To Luminex Multiplex Assays

